**ABSTRACT**

This study was designed to study the effects of stocking density without or with yeast extracts supplementation on productive and reproductive performance, digestive enzymes, blood biochemical parameters, and antioxidant and immune measurements of quail breeders. A total number of 336 Japanese quail birds at 8 weeks old were used. Quails were randomly distributed into 6 groups and 4 replicates maintained / each group in a complete randomized design. The 1st group contained 36 birds (24 females and 12 males) was stocked at a rate of 9 birds/replicate (control, ND; 36 quails/m²) and fed the basal diet without any supplementation; the 2nd, 3rd, 4th, 5th and 6th groups contained 60 quails (40 females and 20 males) and stocked at a rate of 15 chicks/replicate (HD; 60 quails/m²) and fed on a basal diet supplemented with 0, 1, 2, 3 and 4 mg YE/kg diet, respectively. Egg number, egg weight and egg mass were significantly decreased (P<.0001) due to increasing stocking density from ND (9/cage) to HD (15/cage) and fed without dietary YE supplementation. The activity of SOD (p<0.0001), TAC (p<0.0001) and GPX (p=0.0047) was improved with HD plus YE when compared to HD. Digestive enzymes (amylase, lipase, and protease) activity was decreased with HD when compared to ND. It could be concluded that addition of yeast extract can positively mitigate the stress applied to quail raised under high stocking density by enhancing productive and reproductive performance, lipid profile, antioxidants and immunity as well as egg quality criteria.

**Keywords:** yeast, stocking density, performance, blood, quail breeders.

**INTRODUCTION**

Japanese quail is considered one of the important alternative resources of animal protein. Quails have many advantages such as early sexual maturity, small size and high egg production, fast growth, short incubation period, low feed requirements, and less floor space compared with the different species of birds (Padmakumar et al., 2000). Moreover, quails are widely distributed in different countries of the world (Roshdy et al., 2010 and El-Tarabany et al., 2015).

Stocking density increases animals’ welfare but lowering it without guaranteed optimal environmental circumstances is of minimal importance (Umk-Banaš et al., 2014). The optimal housing condition for growing poultry is of considerable interest to researchers, and adequate production conditions are necessary to boost poultry production and improve welfare and profit (Lewko and Gornowicz, 2011; Mesa et al., 2017). Housing conditions can also affect animal welfare and mortality of laying hens (Weimer et al., 2019; Schuck-Paim et al., 2021).

Worldwide quail production has contributed to the poultry sector and will have a bright future due to low production costs and satisfy customer expectations (Mohamed et al., 2019; Abou-Kassem et al., 2019). The objective of the quail producers is to raise the stocking density to obtain further reductions in production costs. However an excessive crowding of chicks might affect performance and wellbeing of chickens maintained in cages (Tayeb et al., 2011; Umk-Banaš et al., 2014). To this regard, no clear advice is provided in the literature on ideal space allowance for quail raising. Stocking stress linked with rising animal density can be alleviated by adding feed supplements with significant antioxidant potential (Attia et al., 2021).

The significance of feed supplements/additives in the chicken sector is quite essential (Bolacali and Irak, 2017; El-Kholy et al., 2019). The proper usage of feed additives can improve feed utilization, production and public health. Probiotics such as yeasts (Saccharomyces cerevisiae; SC) are one of the prominent feed additives and have been used as feed additives for improving animal health and performance (Ogbuewu et al., 2018). Probiotics are live microorganisms that boost birds’ health by competing with bad bacteria, improving the gut balance of microbiota and absorption of nutrients (Al-Khalafiah, 2018; Tomaszewska et al., 2018).

The SC is classified as one of the most yeast species that are introduced to chicken feeds during diet formulations (Duarte et al., 2012). The SC contains high levels of nutrients such as proteins, vitamins and minerals, as well as it contains polysaccharides α-D-mannan, β-D-glucan and chitin (Elghandour et al., 2019) which play a key role in improving microbial balance in poultry intestine (Alizadeh et al., 2016). Therefore, this work was designed to study the effects of stocking density without or with yeast extracts supplementation on productive and reproductive performance, digestive enzymes, blood biochemical parameters, antioxidant and immune measurements and egg quality of laying quail.

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MATERIALS AND METHODS

Experimental birds and management

The present study was carried out at the quail unit, Poultry Department, Faculty of Agriculture, Zagazig University, Egypt. A total number of 336 Japanese quail birds (224 females and 112 males) at 8 weeks old were used. Quails were randomly distributed into 6 experimental groups and 4 replicates maintained / each group in a complete randomized design. The 1st group contained 36 birds (24 females and 12 males) was stocked at a rate of 9 birds / replicate (control, ND; 36 quails/m²) and fed the basal diet without any supplementation; the 2nd, 3rd, 4th, 5th and 6th groups contained 60 quails (40 females and 20 males) and stocked at a rate of 15 chicks / replicate (HD; 60 quails/m²) and fed on a basal diet supplemented with 0, 1, 2, 3 and 4 mg YE / kg diet, respectively, (HD + yeast extract). Quails were housed in wire cages of an open farm during the experiment which started at 8 weeks till 16 weeks of age. Feed and water were available throughout the experiment. All laying hens were subjected to 16 hours of continuous light per day during the laying period. All experimental birds were raised under similar environmental, hygienic and managerial conditions.

Experimental diets

The basal experimental diets were formulated to cover the nutrient requirements of laying Japanese quail as recommended by NRC (1994). The composition and calculated analyses of the experimental basal diets are presented in Table 1.

Table 1. Composition and calculated analysis of the experimental basal diet.

<table>
<thead>
<tr>
<th>Items</th>
<th>(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Yellow Corn (8.5%)</td>
<td>58.70</td>
</tr>
<tr>
<td>Soybean meal (44%)</td>
<td>27.50</td>
</tr>
<tr>
<td>Gluten meal (62%)</td>
<td>4.60</td>
</tr>
<tr>
<td>Soybean oil</td>
<td>1.80</td>
</tr>
<tr>
<td>Limestone</td>
<td>5.54</td>
</tr>
<tr>
<td>Di-calcium phosphate</td>
<td>1.20</td>
</tr>
<tr>
<td>Salt</td>
<td>0.30</td>
</tr>
<tr>
<td>Premix1</td>
<td>0.30</td>
</tr>
<tr>
<td>L-Lysine</td>
<td>0.06</td>
</tr>
<tr>
<td>DL-Methionine</td>
<td>0.00</td>
</tr>
<tr>
<td>Choline chloride (50%)</td>
<td>0.00</td>
</tr>
<tr>
<td>Calculated composition2(%)</td>
<td>2901</td>
</tr>
<tr>
<td>ME, Kcal /Kg</td>
<td>2901</td>
</tr>
<tr>
<td>Crude protein</td>
<td>20.00</td>
</tr>
<tr>
<td>Calcium</td>
<td>2.50</td>
</tr>
<tr>
<td>Nonphytate P</td>
<td>0.35</td>
</tr>
<tr>
<td>Lysine</td>
<td>1.00</td>
</tr>
<tr>
<td>Total sulfur amino acids (M+CY)</td>
<td>0.70</td>
</tr>
</tbody>
</table>

1Layer Vitamin-mineral Premix Each 1Kg contains of vit. A, 3000 IU; vit. D3, 1300 I/CU, vit. E 5 mg; vit. K, 2 mg; vit B1, 0.7 mg; vit. B2, 2 mg; vit. B6, 1.5 mg; vit. B12, 7 mg; Biotin 0.1 mg; Pantothenic acid, 6 g; Niacine, 20 g; Folic acid, 1 mg; manganese, 60 mg; Zinc, 50 mg; Copper, 6 mg; Iodine, 1 mg; Selenium, 0.5 mg; Cobalt, 1 mg.
2Calculated according to NRC (1994).

Egg production

Egg number per hen was calculated by dividing the number of eggs laid by the number of hens for each replicate. Each day, eggs were weighed individually to the nearest 0.01 g for each replicate and the average was calculated. Egg mass was calculated by multiplying egg number by average egg weight.

Feed utilization

Weight of feed consumed (g) per each replicate per period was calculated by subtracting the amount of feed left from that supplied. Regarding feed conversion ratio, the amount of feed (g) required to produce one gram of eggs) was calculated by dividing the amount of feed consumed by egg mass produced.

Egg quality traits

Egg produced in the last 4 days of each period (2week) were collected and then used for egg quality measurements. All egg quality traits (egg shape index, egg shell percent, shell thickness, albumen weight percentage, yolk index, yolk weight percentage, Haugh units, and unit surface shell weight (USSW) (mg/cm²)) were determined according to Romanoff and Romanoff (1949), Yalcin et al., (1990).

Reproductive Performance

At the end of each month, eggs from each treatment were collected and incubated. After hatching, all chicks were counted to compute fertility and hatchability percent, and also non-hatched eggs were broken (Reda et al., 2020).

Biochemical characteristics

Individual blood samples were obtained into EDTA tubes from 4 birds within each treatment (on individual basis) at 5 weeks of age to determine the distinct biochemical features. Each blood sample was spun at 4000 rpm for 15 minutes to separate blood plasma. The colorimetric determination of cholesterol was carried out utilising kit provided by Bio-system according to Allain et al. (1974). (1974). The idea of the approach is that cholesterol forms a colourful complex with acetic anhydride and concentrated sulfuric acid and the coloured complex is measured photometrically. The colorimetric determination of triglycerides was carried out by a particular diagnostic kit provided by Bio diagnostic according to Allain et al. (1974). (1974). After centrifugation, the high-density lipoproteins (HDL), low density lipoproteins (LDL) and the very low-density lipoprotein and the very low-density lipoprotein (VLDL) were determined by enzymatic methods (Myers et al., 1994).

Immunity indices

The levels of immunoglobulin G (IgG) and A (IgA) in the plasma were determined spectrophotometrically using a commercial kit from Biodiagnostic Company (Giza, Egypt) according to (Akiba et al., 1982). The authors determined the level of complement (C₃) in quail plasma, using the ELISA Kit from MyBiosource.com.

Antioxidant parameters

Plasma samples were subjected to the measurement of superoxide dismutase (SOD) and glutathione peroxidase (GPX) activity and total antioxidant capacity (TAC) according Koracevic, et al. (2001). Plasma malondialdehyde (MDA) was determined according to Mihara and Uohiyama, (1978).

Digestive enzymes

Digestive enzymes including the activity of amylase, lipase and protease were determined by the method of Tietz and Fiereck (1966) and Lynn and Clevelett-Radford (1984), respectively.
Statistical analysis

The differences among treatments were statistically analyzed by one-way ANOVA using the SAS General Linear Models Procedure (SAS, 2002) by adopting the following model:

\[ X_{ij} = \mu + T_i + e_{ij} \]

Where: \( X_{ij} \) = An observation, \( \mu \) = Overall mean, \( T_i \) = Effect of treatments \( i = 1, 2, \ldots, 6 \), \( e_{ij} \) = The experimental random error. The significant differences between treatment means were separated by Tukey's Multiple Range-test (P<0.05).

RESULTS AND DISCUSSION

Productive performance (egg number, egg weight and egg mass)

Throughout the interval periods studied (8-12, 12-16 and 8-16 weeks of age) the results in Table 2 showed that, means of egg number, egg weight and egg mass were significantly decreased (P<0.0001) due to increasing stocking density from ND (9/cage) to HD (15/cage) and fed without dietary YE supplementation. Adding YE to diets of HD groups led to increasing the aforementioned traits compared to group housed at normal density and HD without dietary YE supplementation (Table 2).

The results observed productive performance due to stocking density accordance with previous studies (Ratriyanto et al., 2020; El-Sheikh et al., 2016; El-Trabany et al., 2015) who found that, high stocking density decrease egg production, egg weight, egg number and egg mass in Japanese quail. Other observations in these birds (Italian quails) demonstrated that, increasing the stocking density from 38 to 47 quails/m² did not affect their egg production, while increasing the stocking density to more than 56 quail/m² causes a decrease in their egg production (Faitarone et al., 2005). Similarly, Mousavi et al. (2016) and Kang et al. (2016) showed, reducing floor space adversely impacted the laying hens egg production without affecting the egg weight and feed conversion ratio.

The unfavorable effects due to high stocking density may be attributed to the alteration of resting behavior due to disruptions from other birds (Attia et al., 2012). In addition, a high stocking density results in a higher temperature cage due to overcrowding, reduced air quality due to inadequate airflow and increased ammonia (Abudabos et al., 2013; Kang et al., 2016).

The results obtained of YE supplementation in the diet of HD groups agree with Pinar et al. (2013) who reported that, dietary supplementation of 1 % inactivated Yeast in brown layers at 22 weeks old for 14 days increased hen-day production percentages. Likewise, Swain et al. (2011) stated that, dietary yeast supplementation at 1.5 g/kg improved egg weight in Vanarja layer. Moreover, Hassanein and Soliman (2010) demonstrated addition of 0.4 % or 0.8 % yeast culture in layer diets improved egg production.

Contradicting results were obtained by Sanaa (2013) who reported that use of dietary yeast at 0.3- 0.6 % lowered laying rate in birds aged 26 weeks. Also, Dizji and Pirmohammadi (2009) observe that laying hens fed 0.04 % has been reported to lay smaller eggs vs. the control group.

Liu et al., (2002) found an improvement in egg weight for Hy-Line hens (20-wks of age) when fed on diets containing commercial yeast culture product at levels 0.1, 0.2 and 0.3%. Furthermore, Osman (2003) reported that feeding Hy-Line White hens on diets supplemented with the probiotic mixture at levels 1, 2, 3 and 4 g/kg diet resulted in higher egg rate, egg weight and egg mass compared to the control group. Moreover, Kurtoglu et al. (2004) found that supplementation of commercial probiotic at 250, 500 and 750 mg/kg increased production of eggs but decreased the ratio of damaged egg. Diets contain live yeast culture increased egg production and egg weight when turkeys were fed phosphorous-deficient diets (Thayer et al., 1978). In addition, Songsak et al., (2009) reported that supplementation of cassava yeast to 26-wk laying hens had a significant positive effect on egg weight but has negative effect on egg production. Abd EL-Maksoud et al. (2011) observed significant improved in egg production and egg mass with adding 250g Ag diet yeast to local hens.

Abu-Taleb et al. (2006) found that average egg production was significantly increased due to adding yeast by 1% or 1.5% into diets of laying quail when laying Japanese quail was fed on basal diets supplemented. Similar results were obtained by Al-Homidan (2007), who indicated that the addition of dried yeast (SC) to laying rations increased the percentage of egg production traits than control.

Table 2. Egg production of laying Japanese quail as affected by stocking density without or with yeast extract supplementation.

<table>
<thead>
<tr>
<th>Items</th>
<th>ND</th>
<th>HD</th>
<th>HD + Yeast extract level (mg/kg diet)</th>
<th>SEM</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td></td>
</tr>
<tr>
<td>Egg number/bird</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8-12 weeks</td>
<td>23.57b</td>
<td>18.89d</td>
<td>23.01b</td>
<td>20.32&lt;.0001</td>
<td>24.76&lt;.0001</td>
</tr>
<tr>
<td>12-16 weeks</td>
<td>19.44b</td>
<td>15.00d</td>
<td>19.44b</td>
<td>17.04&lt;.0001</td>
<td>26.11&lt;.0001</td>
</tr>
<tr>
<td>8-16 weeks</td>
<td>21.51b</td>
<td>16.94d</td>
<td>21.23&lt;.0001</td>
<td>18.68&lt;.0001</td>
<td>25.44&lt;.0001</td>
</tr>
<tr>
<td>Egg weight (g)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8-12 weeks</td>
<td>12.45b</td>
<td>12.06c</td>
<td>12.27b</td>
<td>12.41&lt;.0001</td>
<td>12.48&lt;.0001</td>
</tr>
<tr>
<td>12-16 weeks</td>
<td>13.44b</td>
<td>13.10c</td>
<td>13.33b</td>
<td>13.33&lt;.0001</td>
<td>13.96&lt;.0001</td>
</tr>
<tr>
<td>8-16 weeks</td>
<td>12.94b</td>
<td>12.58c</td>
<td>12.80b</td>
<td>12.87&lt;.0001</td>
<td>13.22&lt;.0001</td>
</tr>
<tr>
<td>Egg mass (g/bird)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8-12 weeks</td>
<td>291.44b</td>
<td>227.73d</td>
<td>282.29b</td>
<td>252.12&lt;.0001</td>
<td>252.12&lt;.0001</td>
</tr>
<tr>
<td>12-16 weeks</td>
<td>261.22b</td>
<td>196.54d</td>
<td>259.13b</td>
<td>272.01&lt;.0001</td>
<td>364.50&lt;.0001</td>
</tr>
<tr>
<td>8-16 weeks</td>
<td>277.33b</td>
<td>212.13c</td>
<td>270.21b</td>
<td>239.56&lt;.0001</td>
<td>336.72&lt;.0001</td>
</tr>
</tbody>
</table>

Means within the same row with different common superscripts differ significantly.

Feed intake and feed conversion ratio

Quail layers reared in HD without or with dietary YE supplementation at different levels consumed lower feed (P<.0001) as compared with layers reared in ND throughout all the experimental periods studied (Table 3). It is worth to note that, the lowest feed consumed through 12-16 weeks of age was recorded for layers housed at HD with dietary YE supplementation at 2 mg/kg diet. The
Reda, F. M. et al.

reduction in feed consumption may be attributed to reduced feeder space per bird in each cage.

Our results agree with the previous researches (Daniela et al., 2018; EL-Shelih et al., 2016; El-Shafei et al., 2012) who indicated that feed consumption for Japanese quail significantly decreased with increasing cage density. Likewise, the effect of YE on feed consumption agree with Liu et al. (2002) who found reduction feed consumption in quails at 0.2 % yeast. Also, Sanaa (2013) reported that, feed consumption was decreased due to dietary yeast at 3-6 g/kg in laying hens. However, Sehu et al. (1997) noticed similar feed consumption in laying quails fed 1.5 % inactivated brewer’s yeast.

Regarding to feed conversion ratio (FCR), the results in Table (3) showed increasing stocking density of Japanese quail layers from ND (9/cage) to HD (15/cage) without dietary YE supplementation caused a significant inferior FCR values through the interval periods. Statistical analysis indicated that, dietary YE supplementation at different levels of quail layers raised under higher density resulted better (P<0.001) FCR values when compared with quail layers group raised under ND and HD without dietary YE supplementation. It could be noticed that, HD group with dietary 3 mg YE/kg supplementation recorded better FCR values compared with other treatment groups.

The effect of stocking density on FCR values in the present study are in agreement with those of EL-Shelih et al. (2016) who reported that FCR values were significantly poorer with increasing the number of quails/square meter throughout the production period. On the other hand, Santos et al. (2011); EL-Shafei et al. (2012) observed FCR/kg of egg produced was better (P<0.05) for quail layers raised under higher floor space. Moreover, Faitarone et al. (2005) stated that, FCR values of quails were not significantly affected among treatment when stocked at 246, 211, 176 and 151 cm²/quail.

The better FCR values in Japanese quail layers raised under high density due to YE supplementation may be due to increase of egg number and egg weight associated with decreased feed consumption and consequently lead to better FCR values. These results are in agreement with Hassanein and Soliman (2010) who found yeast supplementation at 0.02 - 0.04 % improved nutrient utilization in laying birds. Sehu et al. (1997) observed similar FCR in laying quails fed 15% inactivated brewer’s yeast.

In the same line, Al-Mansour et al. (2011) and Abu-Taleb et al. (2006) found that feed intake was not affected, while feed efficiency was enhanced due to adding yeast to laying quail diets by 1 and 19%. Also, Yalcin et al. (2008) investigated the effect of adding 0.5, 1.0, 1.5 and 2.0 the dried yeast (Saccharomyces cervisiae) to Japanese quail diets on the performance from 1 to 6 weeks old. Results indicated that there was a reduction in feed consumption values of yeast fed groups. Feed conversion ratios improved by 6, 9, 12 and 3% due to adding yeast by levels of 1.5 and 2.0% into diets, respectively.

Table 3. Feed intake and feed conversion ratio of laying Japanese quail as affected by stocking density without or with yeast extract supplementation.

<table>
<thead>
<tr>
<th>Items</th>
<th>ND</th>
<th>HD</th>
<th>HD + Yeast extract level (mg/kg diet)</th>
<th>SEM</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Feed intake (g/bird)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8-12 weeks</td>
<td>968.50a</td>
<td>856.98a</td>
<td>854.17bc 824.67bc 853.83bc 845.25bc</td>
<td>8.124</td>
<td>&lt;.0001</td>
</tr>
<tr>
<td>12-16 weeks</td>
<td>932.50a</td>
<td>825.98a</td>
<td>855.67bc 775.67bc 953.83bc 883.50bc</td>
<td>8.598</td>
<td>&lt;.0001</td>
</tr>
<tr>
<td>8-16 weeks</td>
<td>950.50a</td>
<td>841.48a</td>
<td>854.92bc 800.17bc 903.83bc 864.38bc</td>
<td>6.565</td>
<td>&lt;.0001</td>
</tr>
<tr>
<td>Feed conversion ratio (g feed/g egg)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8-12 weeks</td>
<td>3.30a</td>
<td>3.77a</td>
<td>3.03b 3.27b 2.76d 3.43b</td>
<td>0.044</td>
<td>&lt;.0001</td>
</tr>
<tr>
<td>12-16 weeks</td>
<td>3.64a</td>
<td>4.28a</td>
<td>3.30b 3.52b 2.48d 3.25b</td>
<td>0.042</td>
<td>&lt;.0001</td>
</tr>
<tr>
<td>8-16 weeks</td>
<td>3.43b</td>
<td>3.97b</td>
<td>3.16e 3.34e 2.68e 3.37bc</td>
<td>0.015</td>
<td>&lt;.0001</td>
</tr>
</tbody>
</table>

Means within the same row with different common superscripts differ significantly.

Reproductive performance (Fertility and hatchability percentages)

Data illustrated in Table 4 revealed that, fertility % through 8-12 and 8-16 weeks of age and hatchability % during 8-12 period were significantly (P<0.05) decreased with increasing stocking density without or with dietary YE supplementation when compared with layers stocked at ND. However, fertility % during 12-16 weeks of age and hatchability % through 12-16 and 8-16 weeks of age were not significantly affected due to increasing stocking density with or without dietary YE supplementation.

The decreasing fertility % of eggs produced from quail layers raised at high stocking density may be attributed to the decrease in feed consumption and competition among birds, and consequently lead to decreases in hatchability rate.

The obtained results of fertility and hatchability percentages in the present study agree with EL-Shelih et al. (2016) who observed, the percentages of fertility and hatchability for Japanese quail were increased with increasing cage floor space through 11-21 weeks of age. However, Askar et al. (2012) found that, the percentages of fertility and hatchability for Japanese quails were not significantly affected by stocking density through the period from 10-18 weeks.

Attiia et al., (2021) found that the fertility rate was better (P < 0.01) at the LD than HD. The reduction in fertility percentage at greater density under investigation agrees with Tollba et al. (2006). On the contrary, the non-significant effect of stocking density recorded for hatchability percentages disagrees with the previous authors’ findings. There is little information available in the literature concerning the impact of stocking and its interaction with feed supplement (SC/vit E) on reproductive traits. Therefore, this kind of research is needed to improve animal welfare exposed to stocking stress. Housing stress was reported to influence performance, welfare, antioxidant, and poultry behavior (Narinc et al., 2013).
Table 4. Fertility and hatchability percentages of laying Japanese quail as affected by stocking density without or with yeast extract supplementation.

<table>
<thead>
<tr>
<th>Items</th>
<th>ND</th>
<th>HD</th>
<th>HD + Yeast extract level (mg/kg diet)</th>
<th>SEM</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>Fertility %</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8-12 weeks</td>
<td>89.01b</td>
<td>67.22c</td>
<td>81.52b</td>
<td>82.40b</td>
<td>91.92b</td>
</tr>
<tr>
<td>12-16 weeks</td>
<td>92.22</td>
<td>83.58</td>
<td>92.38</td>
<td>95.45</td>
<td>94.19</td>
</tr>
<tr>
<td>8-16 weeks</td>
<td>90.61a</td>
<td>75.40a</td>
<td>86.96a</td>
<td>88.93a</td>
<td>93.05a</td>
</tr>
<tr>
<td>Hatchability %</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8-12 weeks</td>
<td>78.57a</td>
<td>58.89b</td>
<td>65.55bc</td>
<td>64.78bc</td>
<td>75.28bc</td>
</tr>
<tr>
<td>12-16 weeks</td>
<td>79.11</td>
<td>69.25</td>
<td>79.29</td>
<td>83.07</td>
<td>83.42</td>
</tr>
<tr>
<td>8-16 weeks</td>
<td>78.84</td>
<td>64.07</td>
<td>72.42</td>
<td>73.92</td>
<td>79.35</td>
</tr>
</tbody>
</table>

Means within the same row with different common superscripts differ significantly.

Egg quality traits

The results in Table (5) showed that, increasing the stocking density without or with YE addition in their diets did not significantly affect all quail egg quality traits studied (Albumin, Yolk, Shell thickness, egg shape index, Yolk index, Haugh unit and USSW percentages). These results, agree with Daniel et al. (2018) who found, no significant differences were observed for any of the variables analyzed (specific gravity, egg color, shell thickness and Haugh unit). Also, Lopes et al. (2008) did not find any significant difference of specific gravity, albumin percentage, yolk percentage and bark percentage.

Contradicting results were obtained by Ratrityanto et al. (2020) who stated that, a high stocking density adversely effects egg quality in quail layers, particularly, the egg shell thickness. Likewise, El-Sheikh et al. (2016) reported that, quail layers housed at low density laid eggs had higher shell thickness with or without membranes and egg surface area as compared to those at high density. Moreover, El-Tarabary et al. (2015) indicated that quail layers reared at low density (200 cm² / bird) significantly (P<0.05) increased shell thickness, shell percentage and egg shell weight than those reared at higher density (167 and 143 cm² / bird) at 14 weeks of age.

Regarding the effect of yeast supplementation, Hosseini et al. (2006) observed to significant effect on egg shell thickness, egg shell quality and Haugh unit with using yeast at levels of 0.25, 0.5, 0.75 g/kg diet to layers Maziar et al. (2007) indicated no significant effete on egg quality parameters (shell thickness, shell percentage and Haugh unit) when Hy-Line hens were fed 1 g yeast of SC / kg diet under heat stress 33°C. Songsak et al. (2009) found that supplementation of cassava yeast to 26-wk laying hens had a positive effect on shell thickness, but had no significant effect on albumin weight, yolk weight and Haugh unit. Tollha and EL-Nagar (2008) reported no effect on egg quality in any term by adding Saccharomyces cervisiae to 20-wk local hens reared under high density. Similar result was obtained by Soliman (2002) who added 4g active dried yeast /kg diet to 25-wk Bovans White laying hens. Also, Abd El-Maksoud et al. (2011) observed no effect of egg quality except it improved shell thickness when added 250 g/kg diet dried yeast to local hens.

Table 5. Egg quality traits of laying Japanese quail as affected by stocking density without or with yeast extract supplementation.

<table>
<thead>
<tr>
<th>Items</th>
<th>ND</th>
<th>HD</th>
<th>HD + Yeast extract level (mg/kg diet)</th>
<th>SEM</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>Albumin %</td>
<td>52.75</td>
<td>52.35</td>
<td>53.19</td>
<td>53.07</td>
<td>52.91</td>
</tr>
<tr>
<td>Yolk %</td>
<td>32.09</td>
<td>31.69</td>
<td>32.92</td>
<td>31.75</td>
<td>33.19</td>
</tr>
<tr>
<td>Shell %</td>
<td>15.17</td>
<td>15.98</td>
<td>13.89</td>
<td>15.18</td>
<td>13.89</td>
</tr>
<tr>
<td>Shell thickness</td>
<td>0.23</td>
<td>0.21</td>
<td>0.22</td>
<td>0.21</td>
<td>0.22</td>
</tr>
<tr>
<td>Egg shape index</td>
<td>78.85</td>
<td>78.67</td>
<td>80.57</td>
<td>78.89</td>
<td>79.76</td>
</tr>
<tr>
<td>Yolk index</td>
<td>48.86</td>
<td>47.43</td>
<td>50.1</td>
<td>48.18</td>
<td>50.60</td>
</tr>
<tr>
<td>Haugh unit</td>
<td>83.06</td>
<td>80.75</td>
<td>84.26</td>
<td>81.74</td>
<td>81.22</td>
</tr>
<tr>
<td>USSW</td>
<td>47.67</td>
<td>47.12</td>
<td>47.66</td>
<td>47.62</td>
<td>47.98</td>
</tr>
</tbody>
</table>

Means within the same row with different common superscripts differ significantly.

Digestive enzymes

Digestive enzymes (Amylase, lipase, and protease) activity was decreased with HD when compared to ND (Table 6). Dietary supplementation of YE to HD groups resulted in significant enhances in digestive enzyme comparatively with groups kept at HD fed diet without addition of YE (Table 7). Yeasts including S. cerevisiae play positive roles in improving poultry productivity and public health which promote the efficacy of digestive enzymes (Yoon et al., 2004).

Table 6. Digestive enzymes of laying Japanese quail as affected by stocking density without or with yeast extract supplementation.

<table>
<thead>
<tr>
<th>Items</th>
<th>ND</th>
<th>HD</th>
<th>HD + Yeast extract level (mg/kg diet)</th>
<th>SEM</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>Protease (U/L)</td>
<td>0.51a</td>
<td>0.24b</td>
<td>0.43ab</td>
<td>0.26ab</td>
<td>0.36abc</td>
</tr>
<tr>
<td>Amylase (U/L)</td>
<td>18.80a</td>
<td>10.71c</td>
<td>13.55</td>
<td>24.05b</td>
<td>21.55b</td>
</tr>
<tr>
<td>Lipase (U/L)</td>
<td>8.45a</td>
<td>6.35b</td>
<td>10.30c</td>
<td>11.35c</td>
<td>11.40c</td>
</tr>
</tbody>
</table>

Means within the same row with different common superscripts differ significantly.
Blood constituents

Results in Table (7) show that quails stocked under HD without dietary YE supplementation had higher values of serum ALB (p=0.0118), GLO (p=0.0361), A/B % (p<0.0001), ALT (p=0.0044), AST (p=0.0026), urea (p=0.0251), TC (p=0.0309), TG (p<0.0001), HDL (p=0.0013), and VLDL (p<0.0001) than those stocked under ND and HD with YE supplementation at different levels. However, the data revealed insignificant differences among all treatment groups for serum TP, LDL, creatinine and LDL concentrations. The addition of YE in HD diet alleviated the stress effect induced by high stocking density through decreasing the level of aforementioned parameters and achieved serum blood parameters values equal significantly to quail layers stocked at ND compared with layers stocked at HD without YE supplementation.

The activity of SOD (p<0.0001), TAC (p<0.0001) and GPX (p=0.0047) was improved with HD plus YE when compared to HD (Table 8). While, dietary supplementation of YE to HD groups resulted in significant decreases in MDA (P=0.0155) comparatively with groups kept at HD fed diet without addition of YE. These results are confirmed by Abd El-Wahab et al. (2019) who found that dietary yeast supplementation in Japanese quail increased the activities of antioxidant enzymes (CAT and SOD). On the other hand, Cengiz et al. (2015) observed that no significant differences in blood MDA between low and high stocking densities.

Table 7. Total protein and its fractions, liver, kidney functions and lipid profile of laying Japanese quail as affected by stocking density without or with yeast extract supplementation.

<table>
<thead>
<tr>
<th>Items</th>
<th>ND</th>
<th>HD</th>
<th>HD + Yeast extract level (mg/kg diet)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>TP (g/dL)</td>
<td>3.77</td>
<td>3.59</td>
<td>3.68</td>
</tr>
<tr>
<td>ALB (g/dL)</td>
<td>2.07bc</td>
<td>2.44a</td>
<td>2.06b</td>
</tr>
<tr>
<td>GLOB (g/dL)</td>
<td>1.70a</td>
<td>1.15b</td>
<td>1.63a</td>
</tr>
<tr>
<td>A/G (%)</td>
<td>1.24b</td>
<td>2.18a</td>
<td>1.27b</td>
</tr>
<tr>
<td>ALT (IU/L)</td>
<td>8.12b</td>
<td>16.02a</td>
<td>9.04b</td>
</tr>
<tr>
<td>AST (IU/L)</td>
<td>188.55c</td>
<td>332.31a</td>
<td>286.35ab</td>
</tr>
<tr>
<td>Creatinine (mg/dL)</td>
<td>0.37</td>
<td>0.44</td>
<td>0.33</td>
</tr>
<tr>
<td>Urea (mg/dL)</td>
<td>3.72b</td>
<td>5.84a</td>
<td>3.46b</td>
</tr>
<tr>
<td>Lipid profile:</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>TC (mg/dL)</td>
<td>406.00ab</td>
<td>448.78ab</td>
<td>366.85b</td>
</tr>
<tr>
<td>TG (mg/dL)</td>
<td>162.93bc</td>
<td>232.81a</td>
<td>108.54cd</td>
</tr>
<tr>
<td>HDL (mg/dL)</td>
<td>40.79bc</td>
<td>32.97d</td>
<td>38.90ed</td>
</tr>
<tr>
<td>LDL (mg/dL)</td>
<td>332.62</td>
<td>369.25</td>
<td>306.25</td>
</tr>
<tr>
<td>VLDL (mg/dL)</td>
<td>32.59</td>
<td>46.56a</td>
<td>21.71cd</td>
</tr>
</tbody>
</table>

Means within the same row with different common superscripts differ significantly.

There were no significant differences in lysozyme or Complement 3 activity due to increasing stocking density in quail breeders from ND to HD fed diets supplemented with YE. While, the values of IgG (p=0.0065), and IgA (p=0.0112) were improved with HD plus YE when compared to HD (Table 8). The results of stocking density are supported by Gholami et al. (2020) that stated that broiler chickens reared under high stocking density (20 chicks / m³) had the lowest immune parameters than those reared under 10 and 17 chicks / m² at d 42. Likewise, Heckert et al., (2002) reported that the high stocking density suppression of immunity in broiler chickens. Houshmand et al. (2012) pointed out that broiler chickens subjected to a high stocking density had a lower antibody titer against New castle disease than those subjected to a normal stocking density (Heckert et al., 2002). So, it could be concluded that the high stocking density in the current study resulted in poorer immunity.

Table 8. Antioxidant and immunological indices of laying Japanese quail as affected by stocking density without or with yeast extract supplementation.

<table>
<thead>
<tr>
<th>Items</th>
<th>ND</th>
<th>HD</th>
<th>HD + Yeast extract level (mg/kg diet)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>Antioxidants</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SOD (U/mL)</td>
<td>0.30a</td>
<td>0.13d</td>
<td>0.19cd</td>
</tr>
<tr>
<td>MDA (mmol/mL)</td>
<td>0.45b</td>
<td>0.57a</td>
<td>0.36b</td>
</tr>
<tr>
<td>TAC (ng/ml)</td>
<td>0.35c</td>
<td>0.12a</td>
<td>0.24b</td>
</tr>
<tr>
<td>GPX (ng/ml)</td>
<td>0.31bc</td>
<td>0.20b</td>
<td>0.25b</td>
</tr>
<tr>
<td>Immunity</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IgG (mg/dL)</td>
<td>1.02b</td>
<td>0.42c</td>
<td>0.91bc</td>
</tr>
<tr>
<td>IgA (mg/dL)</td>
<td>0.83ab</td>
<td>0.42c</td>
<td>1.07a</td>
</tr>
<tr>
<td>Lysozyme</td>
<td>0.31</td>
<td>0.15</td>
<td>0.23</td>
</tr>
<tr>
<td>Complement 3</td>
<td>107.55</td>
<td>92.75</td>
<td>105.05</td>
</tr>
</tbody>
</table>

Means within the same row with different common superscripts differ significantly.

SOD: superoxide dismutase; MDA: malondialdehyde; TAC: total antioxidant capacity; GPX: glutathione peroxidase; IgG immunoglobulin G; IgA: immunoglobulin A.

In this respect, El-Sheikh et al., (2016) found significant reductions in plasma total protein fractions of Japanese quail layers by increasing stocking density whereas, a significant increase in total lipids, cholesterol, ALT, plasma total calcium and phosphorous with increasing stocking density. On the other hand, they found...
insignificant differences were observed for serum HDL, LDL and AST levels. Also, Tolba et al. (2006) manifested that, levels of plasma TP, albumin, globulin, A/G%, ALT and AST for Egyptian laying chickens were not affected significantly by different stocking densities. Simsek et al. (2014) displayed that, serum HDL and cholesterol levels were reduced for broiler chicks stocked at lower densities (7.5 bird/m²) when compared with those stocked at higher levels (22.5, 18.75, 15 and 11.5 bird/m²).

Skrbic et al. (2009) reported that, no significant differences were observed of broiler chickens for cholesterol concentrations when stocked at 10, 13 or 16 birds/m². Likewise, El-Deek and Al-Harthi (2004) denoted that, stock density had no significant effect on plasma constituents of broiler chicks, especially ALT and AST activity. While, Erisir and Erisir (2002) manifested that, no significant differences were observed for albumin and total proteins of quails stocked at different stocking density. The improvements of some serum blood constituents due to dietary YE supplementation in the present study agree with many researchers who declared that dietary supplementation of yeast reduced serum cholesterol and triglycerides (Yalcin et al., 2010; Hassanin and Soliman 2010; Yalcin et al., 2014).

CONCLUSION

It could be concluded that use of yeast extract in quail breeder's diets can positively mitigate the stress applied to quail raised under high stocking density by improving productive and reproductive performance, enhancing the antioxidant status, immunological parameters, lipid profile and digestive enzymes.

REFERENCES


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