

EFFECTS OF PREGNANT MARE SERUM GONADOTROPIN ON EGG PRODUCTION AND SOME BLOOD HORMONES OF FAYOUMI CHICKENS

Ali, Nematallah G. M.

Department of Poultry Production, Faculty of Agriculture, Ain Shams University

ABSTRACT

One hundred and twenty Fayoumi pullets at 20 weeks of age were used to study the effect of pregnant mare serum gonadotropin (PMSG) on egg production and some related blood serum hormones. Pullets were randomly divided into four equal groups. Pullets of all groups were housed in floor pens; and food and water were offered *ad libitum*. Birds were fed a diet contained 14% crude protein, 2880 Kcal ME/Kg diet, 3.7% calcium and 0.66% total phosphorous. The experiment started from 20 weeks and extended up to 35 weeks of age.

Birds of group 1 served as control (which were injected with 2ml saline solution/bird) at weeks 20 and 30. Those of groups 2 and 3 were subcutaneously injected with 75 IU of PMSG /bird/day for four consecutive days at weeks 20 and 30, respectively. While those of group 4 were twice injected at weeks 20 and 30.

Obtained results indicate that PMSG injection improved ($P < 0.05$) egg production (number and weight) and feed conversion. Moreover, the injection reduced ($P < 0.05$) the level of triiodothyronine (T_3) in blood serum. While it increased ($P < 0.05$) serum progesterone (P_4), estradiol (E_2), and T_4/T_3 ratio. However, serum thyroxin (T_4) level was not affected by treatment. According to the obtained results it could be concluded that PMSG injection might have a useful effect on egg production of low producer birds when injected slightly before sexual maturity.

Keywords: Pregnant mare serum gonadotropin, egg production, thyroid function, ovarian hormones.

INTRODUCTION

Ovarian follicles are the most important steroid producing structures of the avian ovary. The ovary of a mature chicken generally contains 7-9 large preovulatory follicles arranged in a follicular hierarchy, several postovulatory follicles, and numerous small follicles, which have not entered the follicular hierarchy. It has been reported that in avian preovulatory follicles biosynthesis of sex steroids changes during maturation (Gomez *et al.*, 1998; Lee *et al.*, 1998). The cell theory for steroid production suggested that granular layer of preovulatory follicles primarily produce progesterone that are required as substrate for the production of androgen and estradiol by theca layer (Huang *et al.*, 1979). The recently multiple-cell theory of steroidogenesis suggest that theca layer can also synthesize progesterone, androgen and estradiol independent of granulosa layer (Nitta *et al.*, 1991).

In birds, ovulation and oviposition are processes controlled by LH and sex steroids, including progesterone (P_4). Surges of LH and P_4 have been observed between 4 and 7 h before ovulation in laying chicken (Johnson, 1993). Ovulation of the largest and most mature ovarian follicles occurred 15–30 min after oviposition in turkey hens. (Liu *et al.*, 2001).

Egg production is associated with intensive metabolic activities under the action of gonadotropins (Sturkie, 1986). These hormones affect egg production within physiological limits, to afford behoof metabolic activities for the action of gonadotropins and female steroid hormones. They collectively regulate egg formation from ovulation to oviposition (Armstrong, 1984). Pregnant mare serum gonadotropin (PMSG) is glycoprotein, with two α and β subunits, structurally similar to FSH and LH with higher carbohydrates content, especially sialic acid (Sherwood and McShan, 1977). The higher sialic acid content in PMSG appears to account for its long half-life for several days. Thus, a single injection of PMSG can be of a biological effect, at the target organ for more than a week (Hafez, 1985).

The purpose of the present study was to determine the possibility of accelerating the process of egg formation and thus increasing production rate of Fayoumi chickens by PMSG injection, in accordance with metabolic and steroid hormones.

MATERIALS AND METHODS

Experimental design

This experiment was carried out at private poultry farm near Cairo, Egypt. A total number of 120 female Fayoumi pullets aging 20 weeks with body weight averaged (928 ± 38 gm) was randomly chosen from a large commercial flock. Pullets were randomly assigned to four equal groups. Birds of each group were subdivided into three replicates, with 10 birds each. Birds were housed in a floor pen of 1×1.5 m per replicate and were fed a diet containing 14% crude protein, 2880 Kcal ME/kg diet, 3.7% calcium and 0.66% total phosphorous according to NRC (1994), recommended requirements of low producer strains. The experiment was started from 20 weeks to 35 weeks of age. Feed and water were offered *ad lib.*, while the natural day light was only used. All birds have the same hygienic and mangerial conditions.

Birds' in-group 1 (G1) were untreated and served as control, which were injected at weeks 20 and 30 of age with 2ml saline solution/bird for 4 consecutive days. While those of group 2, 3 and 4 were subcutaneously injected (2ml/bird) of an aquatic solution containing 75 IU of PMSG (Folligon, Intervet international B.V. Boxmeer, Holland) which was immediately dissolved prior to administration. Time of injection differed among groups for 4 consecutive days. Injection of pullets for the second, third and fourth group was applied at 20; 30 and at both 20 and 30 weeks of age, respectively.

Egg production (number and weight) was recorded from the first laid egg up to 35 weeks of age. Age at sexual maturity (SM) was estimated as the day at which the bird laid its first egg.

Blood Sampling

Blood samples were collected between 7:00 to 8:00 O'clock at 20 weeks (before PMSG injection) and at 21, 25, 30, 31 and 35 weeks of age from five randomly chosen birds within each group. About 3ml of blood

were withdrawn from the brachial vein into collecting tubes, immediately centrifuged at 3000 rpm for 15 minutes, serum was then decanted and stored at -20°C till analyses.

Hormonal assays

Direct radioimmunoassay (RIA) technique was performed to determine the serum hormones. Ready antibody coated tube kits of Avian (Diagnostic Product Corporation, Los Angeles) were used according to the procedure outlined by the manufacturer. Serum thyroxine (T_4) and triiodothyronine (T_3) were determined according to May (1978). The T_4/T_3 ratio was then calculated. Serum progesterone (P_4) and estradiol (E_2) were determined according to Etches *et al.* (1981).

Statistical Analysis

Data collected were statistically analyzed by the analysis of variance using the General Linear Model (GLM) procedure of the SAS institute (SAS, 1992). All statements of significance are based on the 0.05 level of probability. Significance of differences among means was tested using Duncan's multiple range test (Duncan, 1955).

RESULTS AND DISCUSSION

1 - Productive performance

Data of productive performance of birds as affected by PMSG injection are listed in Table 1. Obtained data reveal that injected pullets at week 20 of age matured ($P < 0.05$) earlier by about 12 days than controls. Moreover, injected birds at week 20, recorded ($P < 0.05$) higher egg number and ($P < 0.05$) heavier egg weight throughout the experimental period than those of control and those which were injected only at week 30, while the highest egg number and egg weight were recorded for the birds which injected twice at week 20 and 30. The laying intensity could be mainly related to metabolic activity. This observation may also indicate a more complex relationship between thyroid hormones and gonadal function of laying chickens. Since thyroidal hormones are essential for energy metabolism, as reported by Queen *et al.*, (1997); Gomez *et al.*, (1998); Lee *et al.*, (1998) and Liu *et al.*, (2001).

Data of feed intake and feed conversion are listed in Table 1. It seems that injected birds in (G2 and G4) ate ($P < 0.05$) more and converted food to eggs ($P < 0.05$) better than controls. It is possible that nutrient intake increased, in part to meet the additional metabolic requirements of estrogen-sensitive target tissues. This relationship provides an indirect evidence to support the proposal that the increase in nutrient intake following PMSG treatment was required for oviductal tissues development and function for the formation of yolk precursors (Zadworny and Etches, 1988).

2 - Hormonal changes

Thyroid hormones: Data of thyroid activity T_3 , T_4 and T_4/T_3 are presented in Table 2. Obtained data indicate that serum T_3 decreased ($P < 0.05$) due to PMSG injection, while serum T_4 had ($P > 0.05$) unchangeable values. However, T_4/T_3 ratio increased ($P < 0.05$) by PMSG injection.

Table 1: Least squares means \pm SE for productive parameters of Fayoumi chickens as affected by PMSG injection.

Item	Age in weeks	G1	G2	G3	G4
Age at sexual maturity (Days)		159 \pm 2.3 ^a	147 \pm 1.6 ^b	156 \pm 2.1 ^a	147 \pm 1.7 ^b
Egg number/	20 – 25	0.63 \pm 0.1 ^b	0.78 \pm 0.2 ^a	0.66 \pm 0.1 ^b	0.79 \pm 0.2 ^a
Bird / day	25 – 30	0.60 \pm 0.1 ^c	0.75 \pm 0.2 ^a	0.65 \pm 0.2 ^b	0.76 \pm 0.3 ^a
	30 – 35	0.64 \pm 0.1 ^c	0.72 \pm 0.3 ^b	0.77 \pm 0.3 ^a	0.80 \pm 0.3 ^a
Egg weight (gm)	20 – 25	34.2 \pm 1.2 ^b	37.3 \pm 1.2 ^a	34.3 \pm 1.1 ^b	38.2 \pm 1.4 ^a
	25 – 30	37.8 \pm 1.4 ^b	43.6 \pm 1.6 ^a	36.8 \pm 1.0 ^b	43.3 \pm 1.4 ^a
	30 – 35	38.2 \pm 1.5 ^b	43.8 \pm 1.8 ^a	43.3 \pm 1.2 ^b	44.4 \pm 1.7 ^a
Feed intake	20 – 25	88.2 \pm 6.4 ^b	114.1 \pm 7.3 ^a	90.8 \pm 7.1 ^b	118.4 \pm 8.2 ^a
gm/bird/day	25 – 30	93.7 \pm 8.5 ^b	127.6 \pm 10.2 ^a	102.4 \pm 8.3 ^b	124.5 \pm 9.9 ^a
	30 – 35	105.8 \pm 10.0 ^b	132.8 \pm 12.5 ^a	129.8 \pm 10.2 ^b	133.8 \pm 13.8 ^a
Feed conversion	20 – 25	4.1 \pm 0.07 ^a	3.9 \pm 0.02 ^b	4.0 \pm 0.06 ^a	3.9 \pm 0.02 ^b
kg feed/ kg eggs	25 – 30	4.1 \pm 0.06 ^a	3.9 \pm 0.04 ^b	4.2 \pm 0.07 ^a	3.8 \pm 0.04 ^b
	30 – 35	4.3 \pm 0.09 ^a	4.2 \pm 0.07 ^a	3.9 \pm 0.09 ^b	3.8 \pm 0.08 ^b

Least squares means in the same row with no common superscripts differ significantly ($P < 0.05$).

G1: Control. G2: Birds injected by 75 IU of PMSG at week 20 of age.

G3: Birds injected by 75 IU of PMSG at week 30 of age.

G4: Birds injected by 75 IU of PMSG at week 20 and 30 of age.

This is scientifically logic since the ratio is a relation between the concentrations of the two hormones. The results of egg production and thyroid hormones demonstrated that thyroid activity was of essential importance for both initiation and maintenance of egg production. Since it is correlated with the energy metabolism needed for biological endothermic reactions. May (1989) and Queen *et al.*, (1997) speculated that a higher metabolism may caused a rapid conversion from T_4 to T_3 which is considered the more potent thyroidal hormone. Such mechanism could have been responsible for the egg production levels observed in the current study of PMSG injected chickens.

The decline observed in serum T_3 level could also represent other mechanisms being related to feed intake. Sturkie, (1986) and Queen *et al.*, (1997) pointed out that plasma T_3 decreased ($P < 0.05$) by increasing feed intake and speculated that the decrease of T_3 may be also due to the advance of productive status.

Progesterone (P_4) and estradiol (E_2): Obtained data of P_4 and E_2 are shown in Table 3. It was found that both of P_4 and E_2 concentrations in blood serum increased ($P < 0.05$) due to PMSG injection by about 3 times and 5 times for P_4 and E_2 , respectively when compared with controls. The strong positive relationship between gonadotropin and ovarian hormones as reported by Etches (1993) may be strictly indicates the greater increase of P_4 and E_2 concentrations in the blood serum due to PMSG. Tixier-Biochard *et al.*, (1990); Johnson (1993); Lee *et al.* (1998) and Liu *et al.*, (2001) speculated that release of gonadotropins stimulate the ovarian follicles

progesterone secretion. In addition, the variation in serum E₂ level may be a function of existed rate of yolk protein biosynthesis and eggshell calcium deposition (Gruber, 1972). It is well known that laying hens require more calcium during eggshell formation (Goldenberg and Fernandez, 1966). The later authors speculated that an estrogen-induced hypocalcaemia occurs several days before ovulation. Estrogen increases the production of blood-calcium binding proteins. Yolk proteins complexes to calcium are transported to the ovary under the influence of estrogen, (Scheideler and Robeson, 1997).

Table(2): Least squares means ± SE for the serum thyroid hormones of Fayoumi chickens as affected by PMSG injection.

Item	Age in weeks	G1	G2	G3	G4
T ₃ (ng/ml)	20	3.2±0.3 ^a	2.9±0.3 ^a	2.8±0.4 ^a	2.9±0.2 ^a
	21	3.1±0.2 ^a	1.7±0.2 ^b	2.9±0.4 ^a	1.6±0.2 ^b
	25	3.3±0.2 ^a	1.5±0.5 ^b	3.4±0.1 ^a	1.6±0.3 ^b
	30	4.0±0.2 ^a	1.6±0.3 ^b	3.3±0.2 ^a	1.7±0.2 ^b
	31	3.6±0.4 ^a	1.8±0.2 ^b	2.1±0.3 ^b	2.0±0.4 ^b
	35	3.4±0.3 ^a	2.1±0.3 ^b	1.9±0.3 ^b	1.9±0.2 ^b
T ₄ (ng/ml)	20	14.4±0.7 ^a	14.7±0.8 ^a	14.6±0.6 ^a	14.3±0.8 ^a
	21	15.2±0.8 ^a	14.7±0.9 ^a	15.3±0.7 ^a	14.7±0.8 ^a
	25	12.8±0.8 ^a	12.6±0.8 ^a	12.5±0.9 ^a	13.1±0.8 ^a
	30	12.5±0.8 ^a	12.4±0.9 ^a	12.2±0.9 ^a	12.3±0.9 ^a
	31	11.1±0.9 ^a	11.3±0.8 ^a	11.0±0.7 ^a	10.8±0.9 ^a
	35	11.3±0.7 ^a	11.2±0.9 ^a	11.4±0.9 ^a	11.2±0.8 ^a
T ₄ / T ₃	20	4.5±0.9 ^a	5.0±1.0 ^a	5.2±0.8 ^a	5.0±1.1 ^a
	21	4.9±0.8 ^b	8.6±1.6 ^a	5.3±0.9 ^b	9.2±1.7 ^a
	25	3.9±0.9 ^b	8.4±1.2 ^a	3.7±0.8 ^b	8.2±1.1 ^a
	30	3.1±0.7 ^b	7.7±1.4 ^a	3.7±0.7 ^b	7.2±1.3 ^a
	31	3.1±0.8 ^b	6.3±1.1 ^a	5.2±0.9 ^a	5.4±0.8 ^a
	35	3.3±0.7 ^b	5.3±1.0 ^a	6.0±0.9 ^a	5.9±0.9 ^a

Least squares means in the same row with no common superscripts differ significantly (P<0.05).

G1: Control. G2: Birds injected by 75 IU of PMSG at week 20 of age.

G3: Birds injected by 75 IU of PMSG at week 30 of age.

G4: Birds injected by 75 IU of PMSG at week 20 and 30 of age

However, PMSG may create a state of coordination along hypothalamo-hypophyseal-ovarian axis in favor of an efficient egg formation process that resulted in higher egg production in the injected birds. This observation is also supported by the findings of Bedecarrats *et al.* (1997) and Karatzas *et al.* (1997). They speculated that Prolactin (PRL) might act at the neural or pituitary level to inhibit gonadotropin secretion or acts directly on the ovary because gonadotropin-stimulated ovulation and steroid production are inhibited by exogenous PRL. In the present study, PMSG injection showed to activate ovarian function. Obviously, Luteinizing hormone (LH) level is affected by the environmental conditions, especially, photostimulation period.

Alternately, changes in the activity of the hypothalamo-hypophyseal-ovarian axis, due to PMSG injection, luteinizing hormone-releasing factor (LHRF) stimulates the release of LH, which in turn stimulates the secretion of steroids in the ovary (Bedecarrats *et al.*, 1997). These hormonal activities are concomitant with the maturation of the reproductive tract, which leads to augment laying of eggs.

Table(3):Least squares means \pm SE serum progesterone (P_4) and estradiol (E_2), of Fayoumi chickens as affected by PMSG injection.

Item	Age in weeks	G1	G2	G3	G4
P_4 (ng/ml)	20	0.108 \pm 0.01 ^a	0.108 \pm 0.01 ^a	0.107 \pm 0.01 ^a	0.107 \pm 0.01 ^a
	21	0.104 \pm 0.01 ^b	0.356 \pm 0.06 ^a	0.108 \pm 0.01 ^b	0.341 \pm 0.07 ^a
	25	0.117 \pm 0.01 ^b	0.371 \pm 0.04 ^a	0.123 \pm 0.02 ^b	0.372 \pm 0.04 ^a
	30	0.162 \pm 0.02 ^b	0.389 \pm 0.05 ^a	0.168 \pm 0.02 ^b	0.396 \pm 0.06 ^a
	31	0.179 \pm 0.02 ^c	0.380 \pm 0.07 ^b	0.414 \pm 0.06 ^a	0.549 \pm 0.08 ^a
	35	0.191 \pm 0.02 ^c	0.398 \pm 0.07 ^b	0.491 \pm 0.07 ^a	0.538 \pm 0.08 ^a
E_2 (pg/ml)	20	34.7 \pm 2.6 ^a	34.2 \pm 2.3 ^a	34.3 \pm 2.2 ^a	36.5 \pm 2.3 ^a
	21	36.3 \pm 3.4 ^b	208.7 \pm 6.6 ^a	41.6 \pm 2.3 ^b	214.8 \pm 5.7 ^a
	25	41.4 \pm 2.9 ^b	232.5 \pm 4.8 ^a	42.7 \pm 2.8 ^b	233.4 \pm 4.5 ^a
	30	43.8 \pm 3.2 ^b	213.7 \pm 4.3 ^a	53.3 \pm 2.2 ^b	218.5 \pm 4.3 ^a
	31	56.6 \pm 3.1 ^c	228.6 \pm 4.1 ^b	290.9 \pm 5.5 ^a	295.6 \pm 5.4 ^a
	35	72.5 \pm 2.7 ^c	176.9 \pm 3.9 ^b	289.7 \pm 4.6 ^a	294.8 \pm 5.8 ^a

Least squares means in the same row with no common superscripts differ significantly ($P < 0.05$).

G1: Control. G2: Birds injected by 75 IU of PMSG at week 20 of age.

G3: Birds injected by 75 IU of PMSG at week 30 of age.

G4: Birds injected by 75 IU of PMSG at week 20 and 30 of age.

In conclusion, subcutaneous PMSG injection for laying Fayoumi chickens at level of 75 IU/bird/daily for four subsequent days, resulted in significant ($P < 0.05$) increase of egg number, egg weight, feed intake and improved the conversion of food to egg. Moreover, the injection reduced ($P < 0.05$) the blood serum level of T_3 , while it increased ($P < 0.05$) the blood serum concentrations of P_4 , E_2 and T_4/T_3 ratio. However, serum thyroxin (T_4) level was not affected by treatment.

REFERENCES

- Armstrong, D.G. (1984). Ovarian aromatase activity in the domestic fowl (*Gallus domesticus*). J. Endocrinol, 100 : 81-86.
- Bedecarrats, G., D. Guemene and M.A. Richard-Yris(1997). Effects of environmental and social factors on incubation behaviour, endocrinological parameters, and production traits in turkey hens (*Meleagris gallopavo*). Poultry Sci., 76 : 1307-1314.
- Duncan, D.B.(1955). Multiple range and multiple F test. Biometrics, 11 : 1-42.

- Etches, R.J., F., 1993. Symposium: Current advances in reproduction. *Poultry Sci.*, 72:848-849.
- Etches, R.J., F. Croze and C.E. Duke (1981). Plasma concentration of luteinizing hormone, progesterone, testosterone and estradiol in follicular and peripheral venous plasma during the ovulation cycle of the hen. *Recent Adv. Avian Endocrinol*, 33 : 89.
- Goldenberg, H., and A. Fernandez (1966). Simplification method for the estimation of inorganic phosphorus in body fluid. *Clin. Chem.*, 12:871-882.
- Gomez Y., Velazquez P. N., Juarez-Oropeza M. A., Pedernera E.(1998). Steroid metabolism in granulosa and theca interna cells from preovulatory follicles of the domestic fowl (*Gallus domesticus*). *Anim. Reprod. Sci.*, 52: 81-91.
- Gruber, M.(1972). Hormonal control of yolk protein synthesis. In *Egg Formation and Production*. (B.M. Freeman and P.E. Lake ed.) Br. Poultry Sci. Ltd., Edinburgh. Page: 23.
- Hafez, E.S.E.(1985). Endocrinology of reproduction. In *Reproduction in Farm Animals*. Ed. by E.S.E. Hafez. 5th ed. pp. 57-113.
- Huang E. S.-R., Kao K. J., Nalbandov A. V.(1979). Synthesis of sex steroids by cellular components of chicken follicles. *Biol. Reprod.*, 20, 454-461.
- Johnson,A.L.(1993). Regulation of follicle differentiation by gonadotropins and growth factors. *Poultry Sci.*, 72: 867-873.
- Karatzas, C.N., D. Guemene, D. Zadworny and U. Kuhnlein (1997). Changes in expression of the prolactin and growth hormone gene during different reproductive stages in the pituitary gland of turkeys. *Reprod. Nutr. Dev.*, 37: 69-79.
- Lee, K.A., K.K. Volentine and J.M. Bahr(1998). Two steroidogenic pathways present in the chicken ovary: Theca layer prefers Δ^5 pathways and granulosa layer prefers Δ^4 pathways. *Domest. Anim. Endocrinol.*, 15 (1): 1-8.
- Liu, H.K., K.E. Nestor, D.W. Long, and W.L. Bacon. (2001). Frequency of Luteinizing Hormone Surges and Egg Production Rate in Turkey Hens. *Biol. of Rep.*, 64, 1769-1775.
- May, J.D.(1989). The role of the thyroid in avian species. *CRC. Crit. Rev. Poultry. Biol.*, 2: 171-186.
- May, J.D. (1978). A radioimmunoassay for 3,5,3-triiodothyronine in chicken serum. *Poultry Sci.*, 57: 1740-1744.
- Nitta H., Osawa Y., Bahr J.M. (1991). Multiple steroidogenic cell populations in theca of preovulatory follicles of the chicken ovary. *Endocrinology*, 129: 2033-2040.
- NRC (1994). *Nutrient Requirements of Domestic Animals. Nutrient Requirements of Poultry*. 9th Rev. ed. national Academy Press. Washington. DC.
- Queen,W.H., V.L.Christensen and J.D.May (1997). Supplemental thyroid hormones and molting in turkey breeder hens. *Poultry Sci.*, 76: 887-893.
- SAS (1992). *Guide for personal computer*, SAS Institute, Inc. Cary, NC.

- Scheideler, S. F. and L. Robeson (1997). Strain and dietary inclusion rate of oyster shell effects on egg production parameters and shell quality Poultry. Sci. 76:84-92
- Sherwood, O.D. and W.H. McShan (1977). Gonadotrophins, in *Reproduction in Domestic Animals*. ed. by H.H. Cole and P.T. Cupps, 3rd ed. pp. 17-47. NY. Academic Press, USA.
- Sturkie, P.D. (1986). In *Avian Physiology*. 4th ed., (P.D. Sturkie, ed) Spriger-Verlag. New York. Inc.
- Tixier-Biochard, M., J.L. Monvoisin, P. Roembauts and E. Decuyper (1990). Effect of the sex linked dwarf gene on circulating level of 17 β estradiol, progesterone and luteinizing hormone in the laying hen. Br. Poultry Sci., 31: 385.
- Zadworny, D. and R.J. Etches (1988). Effect of pregnant mare serum gonadotropin on plasma prolactin; luteinizing hormone, estradiol and ovarian growth in incubating and out-of-lay turkeys. Poultry Sci., 67: 319-326.

تأثير الحقن بالهرمونات المنشطة للغدد الجنسية (سيرم الفرس الحامل) على إنتاج البيض و بعض هرمونات الدم في الدجاج الفيومي
نعمة الله جمال الدين محمد علي
قسم إنتاج الدواجن - كلية الزراعة - جامعة عين شمس - جمهورية مصر العربية

استخدم في هذه الدراسة عدد ١٢٠ دجاجة فيومي عمر ٢٠ أسبوع متماثلة في الوزن بمتوسط وزن ٩٢٨ \pm ٣٨ جم. قسمت الطيور إلى أربعة مجاميع، تحتوي كل مجموعة على ٣ مكررات. (بكل منها ١٠ طيور) وضعت الطيور في مساكن مفتوحة بمساحة ١,٥م² / ١٠ دجاجات استمرت التجربة من الأسبوع الـ ٢٠ إلى الأسبوع الـ ٣٥ من العمر. غذيت الطيور على علفه تحتوي على ١٤% بروتين خام، ٢٨٨٠ كيلو كالوري طاقة مثيلة/كجم علفه، ٣,٧% كالسيوم و ٠,٦٦% فوسفور كلي.

عوملت المجموعات كالاتي: المجموعة الأولى استخدمت كمجموعة مقارنة حيث تم حقنها بالمحلول الفسيولوجي (٢مل محلول فسيولوجي/طائر يوميا لمدة ٤ أيام متتالية عند الأسبوع الـ ٢٠ و الـ ٣٠) المجموعة الثانية حقنت الطيور بـ PMSG بمستوى ٧٥ وحدة دولية لكل طائر تحت الجلد ولمدة ٤ أيام متتالية في الأسبوع الـ ٢٠ من العمر. طيور المجموعة الثالثة عوملت بالحقن كالمجموعة الثانية ولكن تم الحقن في الأسبوع الـ ٣٠ من العمر. بينما في المجموعة الرابعة حقنت الطيور بنفس المعدل (٧٥ وحدة لمدة ٤ أيام متتالية) في الأسبوع الـ ٢٠ و الـ ٣٠.

أوضحت النتائج أن الحقن زاد معنويا من عدد ووزن البيض وكذلك من كمية الغذاء المستهلك وحسن من معدل تحويل الغذاء إلى بيض. بينما قلل الحقن معنويا من مستوى هرمون T₃ في سيرم الدم وزاد معنويا من مستوى تركيز البروجسترون والاستراديول E₂, P₄ في سيرم الدم. وبصفة عامة فإن الحقن بـ PMSG كان له تأثير إيجابي على زيادة إنتاج البيض في الدجاج الفيومي سواء من ناحية العدد أو الوزن خاصة إذا تم الحقن مرتان عند الأسبوع العشرين ثم الثلاثين.