INFLUENCE OF PROBIOTIC, PREBIOTIC AND/OR YEAST SUPPLEMENTATION IN BROILER DIETS ON THE PRODUCTIVITY, IMMUNE RESPONSE AND SLAUGHTER TRAITS

Riad, Sosan A.¹; H.M. Safaa¹; Fatma R. Mohamed¹; Salwa S. Siam² and Hanan A. El-Minshawy³

- Animal Production Department, Faculty of Agriculture, Cairo University, Giza, Egypt
- ² Poultry Breeding Department, Animal Production Research Institute, Dokki, Giza, Egypt
- ³ Ministry of Agriculture Dokki, Giza, Egypt

ABSTRACT

A total of 630 Arbor Acres broiler chicks one-day old were used to study the effect of probiotic, prebiotic and/or yeast supplementation on the productive performance, immune response and slaughter traits. Chicks were divided randomly into 6 treatments and housed at deep litter in an open house system. Each treatment replicated 3 times (35 chicks per replicate). Treatments were as follows: T1 (control; chicks fed corn-soy basal diet) and in the other treatments diets were supplemented with 1g probiotic/kg diet as Lactobacillus acidophilus (T2), 1g yeast/kg diet as Saccharomyces cervisiae (5x10¹² CFU/g); (T3),1g prebiotic/kg diet as mannanoligosaccharide (T4), 1g probiotic+1g prebiotic/kg diet (T5) or 1g yeast+1g prebiotic/kg die (T6). Results indicated that body weight, body weight gain, feed consumption and feed conversion ratio were improved and mortality rate was decreased in response to dietary biological feed additives. Body weight at 42 days of age was significantly heavier by about 29.5, 21.2, 12.4, 11.3 and 9.9% than control in the T6, T5, T4, T3 and T2, respectively. Moreover, all biological additives caused a significant increment in the count of erythrocytes, leukocytes, lymphocytes, heterophils, H/L ratio and the antibody titer against SRBC's. Dressing, internal organs and immune organs relative weights at 42 days of age were significantly improved by using biological feed additives. For all traits, the best values were obtained in T6 followed by T5. Also, T6 gave the best relative economical efficiency (14.70% more than control group). It could be recommended from this study that supplementation the biological additives to broiler diet from 0 to 42 days of age, as above mentioned, has a positive effect on the productivity, immune response, slaughter traits and the economical efficiency.

Keywords: Probiotic, Prebiotic, Yeast, Broiler Performance, Immune response

INTRODUCTION

Recently, supplementing broiler diets with non-feed additives may be an alternative way to improve protein utilization, digestion and promote broiler performance. The continuous search for maximum feed efficiency in modern poultry production has been considered to be a critical point in broiler rearing. Therefore, probiotics has been used as natural biological non-feed additives which have beneficial effects to poultry by improving its intestinal microbial balance to stimulate the processes of digestion and absorption of nutrients (*Pelicano et al., 2002*).

Practically, the addition of probiotic to broiler diets has been shown to be responsible for improved growth and feed conversion ratio (Kalavathy et al., 2003). Regarding prebiotic, it has been defined as non-digestible feed ingredients, which are growth substrates, especially directed towards potentially beneficial bacteria already existing in caecum and colon. Several studies have shown that the addition of prebiotics to broiler diet improved performance through improving gut microflora balance (Xu et al., 2003 and Pelicano et al., 2004). In respect to yeast, it is a bio-stimulator and immunomodulator which containing live bacterial cultures, that can regulate and optimize the ratios among the different types of micro-organisms in the digestive system, preventing upsets and exerting a stimulating affect on the disintegration and absorption of the nutrient substances (Balevi et al., 2000). Newly, manufactures are producing live yeast (Saccharomyces cervisiae) commercially as growth promoter to avoid the adverse effects of stressful environmental conditions. Results of Santin et al. (2003) revealed that yeast can improve immune activity of birds and reduce the toxic effects of aflatoxin.

The present work was conducted to study the effects of using probiotic, prebiotic and/or yeast as biological feed additives in broiler diets on productive performance, immune response and slaughter traits. Also, evaluating economical efficiency as net revenue per unit of total feed costs was undertaken.

MATERIALS AND METHODS

The present study was carried out in Poultry Research Farm, Animal Production Department, Faculty of Agriculture, Cairo University from April to May, 2009 for 6 weeks life span. A total of 630 Arbor Acres broiler chicks one-day old were used to study the effect of dietary probiotic, yeast and prebiotic on the productive performance, immune response and carcass yield. At one-day old, chicks were wing banded and divided randomly, into 6 equal treatments 105 chicks each in 3 replicates with 35 chicks for each. **Treatments are specified as follows:**

- 1- T1: served as control group and fed corn-soybean basal diet.
- 2- T2: chicks were fed the basal diet as *Lactobacillus acidophilus* (1g/kg diet).
- **3-** T3: chicks were fed the basal diet supplemented with 1g yeast per kg diet as *Saccharomyces cervisiae* (5x10¹² CFU/g).
- **4-** T4: chicks were fed the basal diet supplemented prebiotic as mannanoligosaccharide (1g/kg diet).
- **5-** T5: chicks were fed the basal diet supplemented with 1g probiotic+1g prebiotic per kg diet.
- 6- T6: chicks were fed the basal diet supplemented with 1g yeast+1g prebiotic per kg diet.

All chicks were housed on deep litter in an open house system divided into 18 floor pens (6 treatments x 3 replicates). Birds were fed starter diet (22.27% crude protein and 3058 ME Kcal/kg) during the first 3 weeks then switched to finisher diet (19.20% crude protein and 3172 ME Kcal/ kg) from 3 to 6 weeks of age (Table 1). Birds were exposed to 23 hours daily and reared in environmental conditions. Feed and water were provided *ad-libtum* throught the whole experimental period. Both body weight and feed consumption were recorded biweekly and mortality was recorded daily.

Three chicks per each replicate were injected at 35 and 42 days of age intravenously with 1 ml of 7% suspension of sheep red blood cells (SRBC'S). Seven days later, blood samples were collected and centrifuged. Sera were frozen until the measurements of the antibody titer against SRBC'S were determined as humoral immune response according to *Van der Zijpp et al.* (1983) and Bachman and Mashaly (1986). At 42 days of age, 3 chicks per replicate were randomly chosen weight, slaughtered and eviscerated. Internal and immune organs were weighted and carcass traits were measured.

After slaughtering, fresh blood samples were collected to determine the hematological picture for counting the red blood cells (RBC's) and white blood cells (WBC's) according to *Wintroba (1967)*. Lymphocytes and heterophils were counted according to *Haddad and Mashaly (1990)*. Also, small intestine samples were collected to count, examine and define their bacterial content using the procedure of *A.O.A.C. (1995)*.

Table (1): Composition and	d calculated	analyses of	basic diets.
----------------------------	--------------	-------------	--------------

Ingredients	Starter diet (%)	Finisher diet (%)		
Yellow corn	52.03	61.38		
Soya bean meal (44%)	29.60	22.50		
Corn gluten meal (60%)	7.00	6.10		
Vegetable oil	4.00	4.00		
Wheat bran	2.80	2.60		
Bone meal	3.30	2.15		
Limestone	0.14	0.14		
Premix	0.30	0.30		
NaCI (salt)	0.50	0.50		
L-lysine-HCL	0.18	0.18		
DL- Methionine	0.15	0.15		
Total	100	100		
Calculated values				
Crude protein (%)	22.27	19.20		
ME (Kcal/Kg)	3058	3172		
Crude fiber (%)	3.80	3.32		
Ether extract (%)	6.50	6.73		
Calcium (%)	0.92	0.73		
Available P (%)	0.48	0.40		
Lysine (%)	1.22	1.05		
Methionine (%)	0.55	0.49		
Methionine + Cystine (%)	0.92	0.81		

Supplied per Kg diet: Vit.A, 12000 IU; vit D3, 2000.000 IU; Vit.E, 10 mg; Vit.K, 2 mg; Vit.B₁, 1mg; Vit.B₂, 5mg; Vit.B₆, 1.5 mg; Vit.B₁₂, 10 μ g; Nicotinic acid 30mg; Folic acid 1mg; Pantothenic acid 10mg; Biotin 50 μ g; Choline chloride 500mg; Copper 10mg; Iron 30mg; Manganese 60mg; Zinc 50mg; Iodine 1mg; Selenium 0.1mg and cobalt 0.1mg.

According to NRC (1994).

Economical efficiency of production was calculated from the inputoutput analysis of the money, based on the differences in both growth rate and feeding costs. The value of the economical efficiency was calculated as net revenue per unit of total feed costs. The prices of experimental diets and live body weight were calculated according to the prices of the local Egyptian market at the time of experiment.

All results were statistically analyzed as one-way analysis of variance by using the General Linear Model (GLM) adapted to micro computer of statistical analysis system (SAS, 1999). Differences between means were tested by multiple range tests according to *Duncan* (1955).

RESULTS AND DISCUSSION

I. Productive performance traits:

a. Live body weight and body weight gain:

Results in (Table, 2) indicated that the addition of the tested biological additives in broiler diet from 0 to 42 days of age improved both body weight and body weight gain during the experimental period compared to the control. Weight at 42 days of age was significantly heavier by about 29.5, 21.2, 12.4, 11.3 and 9.9% than control in the T6, T5, T4, T3 and T2, respectively. Body weight gain from 0 to 42 days of age exhibited also the same trend. The best values of both traits were recorded by using yeast+prebiotic (T6) followed by probiotic+prebiotic in diet (T5), then prebiotic (T4), yeast (T3) and probiotic (T2). An improvement in productive performance were obtained also by using Lactobacillus (Dilworth and Day, 1978 and Jernigan et al., 1984), Lacto-sacc (Gippert and Bodrogi, 1992 and Ali, 1994). Yea-sacc and Fermacto (Omar. 1996), Yoghurt (El-Deeb and Makled, 1993), butter milk (Ghazalah and Ibrahim, 1998) for broiler diets and yeast culture (Ali et al,. 2000) for Japanese quail diets. Lactic acid bacteria help to maintain an optimum low pH to inhibit growth of undesirable bacteria (Alltech Bio-Technology Center Announcement, 1989).

Ali (1999) obtained 5.1% improvement in live body weight at 7 weeks of age when broiler chicks were fed Aspergillus supplemented diet. The improving of probiotic was explained by the favorable effects that provide live yeast culture and natural lactic acid producing bacteria to the chick's digestive tract, where the live yeast culture stimulates fiber digestion to guard against digestive upset. Santin et al. (2001) reported a significant increase in weight gain when birds fed 0.2% cellular wall from Saccharomyces cervisiae (Mannan-oligosaccharide). Similar result was found by El-Sheikh et al. (2008) who used manna-oligosaccharides (MOS) at 21 days of age. Vytautas et al. (2006) explained that the positive results of prebiotic on body weight and weight gain may be attributed to that additive may help to maintain the microflora balance of the intestinal tract of chicken resulting in a more efficient use of nutrients from feed, more intensive processes of protein metabolism and subsequently in better health.

b. Feed consumption and feed conversion:

Data in (Table, 3) indicated that feed intake was significantly higher in T5, T6, T4, T3 and T2 than in T1 by 13.71, 13.43, 11.38, 8.52 and 3.09%, respectively. Also, feed conversion from (0-42 days of age) was improved, in which the best values were in order for T6, T5, T2, T3 and T4 compared to the control one. In agreement with our results, Madkour et al. (2008) reported that using probiotic and prebiotic in broiler diet improved significantly both feed consumption and feed conversion from 0-42 days of age. March (1979) suggested that these improvement by probiotic may be due to that intestinal pH may alter both microbial population and nutrient absorption and this may improve efficiency of feed utilization. Also, Sellars (1991) reported that probiotic presence high numbers of Lactobacilli that increase the motility of gut content and improve nutrient availability or absorption which leads to improve efficiency of feed utilization. Haddadin et al. (1996) reported that the change of microbial in bird's intestine might enhance their health and improve feed consumption by using probiotics. Moreover, Tomasik and Tomasik (2003) and Kirkpinar et al. (2004) stated that prebiotics belong to a group of indigestible dietary carbohydrates improved feed conversion. Also, EL-Nagmy et al. (2007) stated that probiotic addition in broiler diet lead to improve growth rate. In addition, recently, Tollba and Mahmoud (2009) found that feed intake increased by 5.32% at normal temperature (23°C) and by 7.97% at high temperature (38°C), when chicks fed diets with probiotic (biogen) at 2g/kg of feed. Similar results were obtained by Kalavathy et al. (2003) and Tollba et al. (2004).

c. Mortality rate:

Cumulative mortality rate from 0 to 42 days of age is presented in Table (4). Results indicated a depression in mortality rate to 3.81, 10.46, 8.76, 6.49 or 5.56% in T6, T5, T4, T3 or T2 compared to 13.27% in control one, respectively. The result was agreed with those reported by *Hussein and El-Ashry (1991), Rashwan et al. (1993), Tawfeek et al. (1993) Omar (1996) and Alm Eldein (2002)* who stated the addition of probiotic decreased mortality rate. *Watkins and Miller (1983)* attributed the reduction of mortality rate in *Lactobacillus acidophilus* treatments to the inhibitory effects of probiotic towards enteric micro-organisms. This effect was due to the alteration of pH through acid production, and changes the oxidation-reduction potential through its production of metabolites, by making the environment less conductive for organisms requiring oxygen (*Hamdy et al., 2009*). *Abd El-Samee (2001)* reported that the common pathogenic bacteria like *E.coli, Salmonella and Clostradia* which caused bacterial diseases such as diarrhea, enteritis and/or mortality could be prevented when probiotic was used.

II. Immunological traits:

a. Blood hematological picture:

Results in (Table, 5) declared significant increase in counts of erythrocytes, leukocytes, lymphocytes, heterophils and H/L ratio in biological additives treatments than control ones. However, values were better in T6, T5, T4, T3 than T2.

2-3

4-5

The result agrees with that reported by *Zulkifli et al. (2000)* who stated an enhancement of immunity that might be expected corresponding to adding probiotic.

Tollba and El-Nagar (2008) obtained significant increase in counts of erythrocytes, leukocytes, lymphocytes due to feeding dietary biogen (bacteria concentration as probiotic) compared with control diet. Tollba and Mahmoud (2009) reported a significant increase in counts of erythocytes, leukocytes, lymphocytes, eosinophils and basophils, but not in heterophils, by feeding ditary biogen at normal and high temperature compared to the control ones.

b. Humoral immune response:

The same trend was observed also for immune response that presented in Table (5). Significantly high primary and secondary immune response which represented as the antibody titer against SRBC's at 35 and 42 days of age were found in biological additive treatments than control ones. In general, T6 was better than T5, T4, T3 and T2. Similar result was reported by Malzone et al. (2000) and Shashidhara and Devegowda (2003) who observed a significant increase in the antibody titers against SRBC's for birds fed 0.05% MOS compared to control ones. Similarly, Watkins et al. (1982) and Zulkifli et al. (2000) stated that the immune response of broilers was increased when chicks fed diet containing Lactobacillus cultures. The effect of antibody titers may be due to the influence of the MOS on immune system and/or the improvement occurred in the intestinal absorption for nutrients, such as: Zn, Cu and Se. In addition, it might be attributed to the reduction of the pathogenic bacteria load in the intestine which preventing the acute immune response against such bacteria (Finucane, et al., 1999 and Spring et al., 2000). Ferket et al. (2002) showed that the immune modulation could also be improved by stimulating gut-associated and systemic immunity as a nonpathogenic antigen providing an adjuvant like effect.

On the other hand, *Al-Homidan and Fahmy (2007)* did not indicate significant effect of yeast culture on antibody titer against SRBC's in broilers. Also, *Hassan and Ragab (2007)* reported insignificant increase in primary and secondary antibody titer against SRBC's of laying hens fed MOS in diet.

III. Bacteria enumeration and microbial status:

As shown in (Table, 6) adding the biological additives improved significantly (P \leq 0.05) the number of the intestinal microbial. So lactobacillus number in T2 and T5 as well as yeast amount in T3 and T6 were significantly increased than control ones. It can be summed, that using biological additives caused beneficial effects that improve intestinal microbial balance, as well as stimulated growth processes and improved productive performance (Tables 2 and 3) and decreased mortality rate (Table 4). In this connection, *Kamra and Pathak (1996)* reported that yeast culture (*Saccharomyces cervisiae*) improved the digestibility of crude protein and crude fiber fractions, thereby increasing the availability of nutrients for animal productively (*Kruase et al., 1989 and Bradley et al., 1994*) or indirectly via change in the gut microflora in favour of the activities of fiber degrading micro-organisms especially

J. of Animal and Poultry Production, Vol. 1 (2), February, 2010

cellulolytic bactria and subsequently decreasing non-starch polysaccharides contents (NSP's) in the gut (*Miles, 1993*). Furthermore, probiotics produce lactic acid which alter the pH of chicken gut making it improper media for harmful bacteria such as salmonella and pathogenic species of *E. coli* (*Leesson and Major, 1990*), improve nutrient availability and absorption (*Sellars, 1991*), stimulate appetite (*Nahashon et al, 1994*), produce digestive enzymes (*Lee and Lee, 1990*), and improve intestinal microbial balance (*Fuller, 1989*).

fr	from 0 to 42 days of age.									
	Experimental treatments									
Traits Items Control Probiotic Yeast Prebiotic Probiotic + Yeast + Prebiotic Prebiotic										
Total count	2.14 ^b	18.00 ^a	9.63 ^{ab}	9.10 ^{ab}	4.64 ^b	6.53 ^b				
(x10 ⁶)	±0.82	±2.31	±1.77	±1.02	±1.86	±2.79				
Yeast (x10 ³)	5.93 ^b		59.00 ^a			27.20 ^{ab}				
reast (XTU)	±1.93	-	±4.73	-	-	±19.63				
Lactobacillus	5.87 ^b	406.67 ^a			7.70 ^b					
(x10 ³)	±1.17	±93.51	-	-	±0.57	-				

Table (6): Counts of some intestine pathogenic bacteria of Arbor	Acres
broiler chicks fed diets with different biological ad	ditives
from 0 to 42 days of age.	

^{a,b} Means within rows with different superscripts are significantly different (P≤0.05).

IV. Carcass traits:

Values in (Table, 7) stated that all studied traits (except abdominal fat) were significantly higher in chicks fed biological additives than control ones. However, values were higher in T6, T5, T4, T3 than T2. Similar results were reported by *Rashwan et al. (1993), EL-Gendi et al. (2000) and Madkour et al. (2008)* whom obtained significantly improved values for dressing%, internal and immune organs at 6 weeks of age in broilers fed biological probiotic and prebiotic than control ones. Moreover, probiotic treatments improved carcass weight (*Alm Eldein, 2002*) and carcass % (*Yusrizal and Chen, 2003*).On the other hand, *Ali (1999)* reported insignificant improvement in carcass weight, total edible parts for broilers fed probiotic diets compared with control.

Concerning, abdominal fat (Table 7), it decreased by about 36.1, 33.1, 29.0, 26.6 and 24.9% in T6, T5, T4, T3, T2 than control ones, respectively. This result agrees with *Ali (1999) and Yusrizal and Chen (2003)* who found that abdominal fat reduced with biological additives.

V. Economic efficiency:

Data in (Table, 8) indicated that both net revenue and economical efficiency increased in biological additives treatments than control ones. Relative economical efficiency% showed an improvement by about 14.70, 11.18, 7.62, 1.26, and 0.25% in T6, T3, T2, T5 and T4 than control, respectively. The result agrees with *Namra (2006)* who suggested that incorporation of 0.15 % baker's yeast in diet of quail layers apparently exhibited better amelioration in feed cost /egg than the control group.

7-8

Hassan and Ragab (2007) stated that MOS supplementation into layer diets improved economical efficiency and relative economical efficiency of laying hens than those fed the control diet. *Tollba and Mahmoud (2009)* indicated that addition of probiotic (biogen) at 2g/ kg of broiler diets increased significantly economical efficiency by 26.13% at normal temperature (23°C) and 3.74% at high temperature (38°C).

It could be concluded that the biological additives to broiler diet improved all productive traits, immune response and carcass yield. For all studied traits, the best values were obtained for chicks fed on yeast + Prebiotic diet. Also, it gave the best relative economical efficiency%.

REFERENCES

- Abd El-Samee, M.O. (2001). Broiler performance as affected by crude protein, lysine and a probiotic. Egypt. Poult. Sci., 21 (4): 943-962.
- Alm Eldein, A.K. (2002). A comparative study on reducing the cholesterol in eggs of laying hens. Ph.D. Thesis. Fac. Agric. Cairo Univ., Egypt.
- Al-Homidan, A. and M.O. Fahmy (2007). The effect of dried yeast (Saccharomyces cervisiae) supplementation on growth performance, carcass chemical analysis, immunity, ileum vili hights and bacterial counts of broiler chickens. Egypt Poult. Sci., 27 (III): 613-623.
- Ali, A.M., K.Y. El-Nagmy and M.O. Abd El-Samee (2000). The effect of dietary protein and yeast culture levels on performance of growing Japanese Quail. Egypt. Poult. Sci., 20: 777-787.
- Ali, M.A. (1994). The effect of some nutritional additives on feeding broiler. M.Sc. Thesis. Faculty of Agriculture, Zagazig University.
- Ali, M.A. (1999). Effect of probiotics addition to broiler nation on performance and some blood constituents. Egypt. Poult. Sci., 19:161-170.
- Alltech Bio-Technology Center Announcement (1989). All Natural Rabbit Program 3031 Catrip Hill Pike Nicholasville, K.Y. 40336, U.S.A.
- A.O.A.C. (1995). Association of Official Analytical Chemists. Official Methods of Analysis, 16th ed.Washington, DC.
- Bachman, S.E. and M.M. Mashaly (1986). *Relationship between circulating thyroid hormones and inmature male chickens. Develop Comparat Immunol.*, 10: 395-403.
- Balevi, T., S. Uean, B. Coskun, V. Kurtoglu and S. Cetingul. (2000). Effect of commercial probiotics in the diet on performance and immune system in layers. Hayvancilik-Arastirma-Dergisi., 10: 1-2, 25-30.
- Bradley, G.L., T.F. Savage and K.I. Timm (1994). Effect of supplementing diets with Saccharomyces cervisiae var. boulardii on male poultry performance and ileal morphology. Poult. Sci., 73: 1766-1770.
- Dilworth, B.C. and E.J. Day (1978). Lactobacillus cultures in broiler diets. Poult. Sci., 57: 1101 (Abst.).
- Duncan, D.B. (1955). Multiple range and multiple F tests. Biometrics 11: 1-42.
- El-Deeb, M.A. and M.N. Makled (1993). Preliminary study on the effect of yoghurt as natural probiotic supplement on some growth parameters of broiler chicks. Egypt. Poult. Sci., 13: 411-427.

- EL-Gendi, G.M., A. F. Soliman and A. G. Habib (2000). Evaluating four feed additives for improving productive and metabolic performance of broiler chicks. Egypt. Poult. Sci., 20: 103-122.
- EL-Nagmy, K.Y., A.A. Ghazalah and A.S. Bahakim (2007). The effect of probiotics supplementation on performance of broiler chicks fed diets varying in protein content. 4th World Poultry Conference, 27-30, Sharm El-Sheikh, Egypt.
- El-Sheikh, T.M., M.O. Abd El-Samee and A.M. Ali (2008). Effect of reusing broiler litter and Bio-Mos as alternative of growth promoters on broiler performance during summer season. Egypt. Poult. Sci., 28(I): 265-282.
- Ferket, P.R., C.W. Paks and J.L. Grimes (2002). Benefits of dietary antibiotic and mannan oligosaccharide supplementation for poultry. Page 5-24 in Proceedings of the Multi-State Poultry Feeding and Nutrition Conference. Indianapolis, IN.
- Finucane, M., P. Spring and E. Newman (1999). Incidence of mannose sensitive adhesions in enteric bacteria . Poult. Sci., 78 (Suppl.): 139 (Abst.).
- Fuller, R. (1989). Probiotict in man and animal. A review J. Appl. Bacter., 66:365-378.
- Ghazalah, A.A. and F.A.A. Ibrahim (1998). Effects of dietary supplementation of butter milk as natural probiotic on the performance of growing and laying Japanese quails. Egypt. Poult. Sci., 18 (1): 61-80.
- Gippert, T. and G. Bodrogi (1992). *Lacto-Sacc in the feeding of broiler duck. Proceed, 9th Int. Sump. On waterfowl, pp. 16-18, Pisa, Italy.*
- Haddad, E.F. and M.M. Mashaly (1990). Effect of thyrotropin releasing hormone, triiodothyronine and chicken growth hormone on plasma concentration of throxine, triiodothyronine, growth hormone and growth of lymphoid organs and leukocyte populations in immature male chicken. Poult. Sci., 69: 1094-1102.
- Haddadin, M.S.Y., S.M. Abdulrahim, E.A.R. Hashlamoun and N.K. Rabinson (1996). The effect of Lactobacillus acidophlus on the production and chemical composition of hen's egg. Poult.Sci., 75: 491-494.
- Hamdy, A.M.M; G. Tellez and B.M. Hargis (2009). Why is Lactobacillus acidophilus so important for chicken's health. Egyptian J. Nutrition and feeds, 12 (3), Special Issue; 761-769.
- Hassan, H.A. and M.S. Ragab (2007). Single and combined effects on mannanoligosaccharide (MOS) and dietary protein on the performance and immune response of laying hens. Egypt Poult. Sci., 27 (IV): 969-987.
- Hussein, H.H. and M.A. El-Ashry (1991). Some studies on the beneficial effects of Lactobacillus concentrate supplementation on broiler performance. Egypt. J. Anim. Prod., 28 (1): 85-91.
- Jernigan, M.A., R.D. Milles and A.S. Arafa (1984). Probiotic in poultry nutrition: A Review. World's Poult. Sci., 12: 99-107.

- Kalavathy, R., N. Abdullah, S. Jalaludin and Y.W.Ho (2003). Effect of Lactobacillus cultures on growth performance, abdominal fat deposition, serum lipids and weight of organs of broiler chickens. Br. Poult. Sci., 44: 139-144.
- Kamra, D.N. and N.N. Pathak (1996). *Feed additives. Pages 170-178 in: Nutritional Microbiology of Farm Animals.* 1th Ed. Vikas Publishing house PVT.LTD. New Delhi, Indian.
- Kirkpinar, F., Z. Acikgon and V. B. Ayhan (2004). Effects of inclusion of poultry by product meal and enzyme probiotic supplementation in grower diets on performance and feed digestibility of broilers. Br. Poult. Sci., 45: 273-279.
- Kruase, O.G., C.R. Richardson, R.E. Castleberry and C.W. Cobb (1989). Biological responses of chicks fed sorgum grain based diets with added grain specific enzymes mixture and yeast, 1989. Texas tech of Agriculture Science, Lubbock. T5. 236: 7-8.
- Lee S.Y. and B.H. Lee, (1990). Esterolic and lipolytic activities of lactobacillus casi-subspcasei LLG. J. of Food Sci., 55: 119-122
- Leesson, S. and D. Major (1990). As biotechnology grains momentum Canadian researchers study need for feed criteria. Feed Stuffs. 62: 23 – 30.
- Madkour, M.A., I. El-Wardany, S.A. Yassein, H.M.H. El-Allawy and A.A. Mekhaimar (2008). *Performance of broiler chicks as influenced by adding some biological and natural growth promoters. Egypt. Poult. Sci., 28(I): 83-102.*
- Malzone, A., B. Paluch, M.S. Lilburn and A.E. Sefton (2000). Modulation of humoral immunity in commercial laying hens by dietary probiotic. Poult. Sci., 79 (suppl 1), 165. (Abst).
- March, B.E. (1979). The host and it's microflora: An ecological Unit. Journal Animal Science., 49: 85-89.
- Miles, R.D. (1993). Manipulation of microflora of the gastrointestinal tract: Natural ways to prevent colonization by pathogens pages 133-150. In Biotechnology in the feed industry. Proc. Alltch Ninth Animal Symposium T.P. Lyons, ED. Alltch technical Publications, Nicholasville, KY, USA.
- Nahashon, S.N., H.S. Nakaue and L.W. Mirosh (1994). Production variables and nutrient retention in Single Comb White Leghorn laying pullets fed diet supplemented with direct fed microbial. Poult. Sci., 73: 1699-1711.
- Namra, M.M.M. (2006). Influence of using Baker's Yeast and microbial phytase in Japanese Quail diets on productive performance and some physiological parameters. Egypt. Poult. Sci., 26: 579 – 607.
- NRC, (1994). National Research Council. Nutrient requirements of poultry. 9th Ed. National Academy Press, Washington, DC.
- Omar, A.S. (1996). Bio protein in feeding broiler chicks. M.Sc. Thesis. Faculty of Agriculture, Zagazig University.
- Pelicano, E.R.L., P.A. Souza and H.B.A. Souza (2002). Prebiotics e probiotics na nutricao de aves. Ciencias Agrarias e da Saude. 2: 59-64.

- Pelicano, E.R.L., P.A. Souza and H.B.A. Souza, A. Oba, K.A. Norkus, L.M. Kodawara and T.M.A. Lima (2004). Performance of broilers fed diets containing natural growth promoters. Revista. Brasileira. de. Ciencia. Avicola, 6: 231-236.
- Rashwan, A.A., K.A. Yamani, M.I. Tawfeek and S.S. Ahmed (1993). Effect of genotype and Fermacto supplementation to the commercial diet on some economic production traits in broiler chickens. J. Agric. Sci., Mans. Univ., 18 (8): 2270-2278.
- Santin, E., A. Maiorka, M. Macari, M. Grecco, J.C. Sanchez, T.M. Okada and A.M. Myasaka (2001). *Performance and intestinal mucosa development of broiler chickens fed diets containing Saccharomyces cervisiae cell wall. J. Appl. Poult. Res., 10: 236-244.*
- Santin, E., A.C. Paulilo, A. Maiorka, L.S.O. Nakaghi, M. Macari, A.V.F. Silva and A.C. Alessi (2003). Evaluation of the efficacy of Saccharomyces cervisiae cell wall to ameliorate the toxic effects of aflatoxin in broilers. Int. J. Poult. Sci., 2(5): 341-344.
- SAS. (1999). SAS User's Guide. Statistics. Version 8, Edition SAS Inst., Inc., Cary. NC.
- Sellars, R.L. (1991). Acidophilus producuts: Therapeutic probiotics of fermented milks. R.K. Robinson, ed. Chapman and Hall, London, UK. pp: 81-116.
- Shashidhara, R. G. and G. Devegowda (2003). Effect of dietary mannanoligosaccharide on broiler breeder production traits and immunity. Poult. Sci., 82:1319(Abst.).
- Spring, P., C. Wenk, K.A. Dawson, and K.E. Newman (2000). The effect of dietary mannan-oligosaccharides on cecal parameters and the concentrations of enteric bacteria in the caeca of Salmonella-challenged broiler chicks. Poult. Sci., 79, 205-211.
- Tawfeek, M.I., K.A. Yamani, A.A. Rashwan and S.S. Ahmed (1993). Growth performance, carcass traits and blood constituents in broiler chicks as affected by genotype: Dietary protein sources and feed additions. J. Agric. Sci., Mans. Univ., 18 (8): 2279-2289.
- Tollba, A. A. H. and A.H.I. El-Nagar (2008). Increasing stocking density of Egyptian laying hens by using: 1- Antioxidants & live yeast. Egypt. Poult. Sci., 28 (II): 575-598.
- Tollba, A.A.H. and R.M. Mahmoud (2009). How to control the broiler pathogenic intestinal flora under normal or heat stress conditions. 1-Medical plant- Probiotics- Sand as a litter. Egypt. Poult. Sci., 29 (II): 565-587.
- Tollba, A.A.H., M.M. Sabry and S.M.M. Abuzead (2004). Effect of microbial probiotics on performance of broiler chicks under normal or heat stress conditions: 1- Lactobacilus or Pediococus. Egypt. Poult. Sci., 24 (II):351-367.
- Tomasik, P.J. and P. Tomasik (2003). *Probiotics and prebiotics. Cereal Chem.*, 80: 113-117.

- Van der Zijpp, A.J., K. Frankena, J. Boneschancher and M.G.B. Nieuwland (1983). Genetic analysis of primary and secondary immune response in the chicken. Poult. Sci., 62: 565-572.
- Vytautas, S., R. Bobiniene, D. Gudavicue, R.Cepuliene, V. Semaska, D.Vencius and I. Kepaliene (2006). *Influence of a feed additive on* some biochemical indices of blood and intestinal microbial of broiler chickens. Lietuvos, Mokslu, Akademija. 4, 57-62.
- Watkins, B.A. and B.F. Miller (1983). Competitive gut exclusion of avian pathogens by Lactobacillus acidophils in ganotobiotic chicks. Poult. Sci., 62: 1772-1779.
- Watkins, B.A., B.F. Miller, D.H. Neil and M.T. Collins (1982). In vivo inhibitory effects of Lactobacillus acidophils against pathogenic E. coli in gnotobiotic chicks. Poult. Sci., 61: 1298-1308.
- Wintroba, M.M. (1967). Clinical Hematology. 6th Edition PP., 414:419, Lea Febiger, Philadelphia, U.S.A.
- Xu, Z.R., C.H. Hu, M.S. Xia, X.A. Zhan and M.Q. Wang (2003). Effect of dietary fructo oligosaccharide on digestive enzyme activities. Intestinal Microflora and morphology of male broilers. J. Anim.Sci., 82: 1030-1036.
- Yusrizal and T.C. Chen (2003). Effect of adding chicory fructans in feed on broiler growth performance, serum cholesterol and intestinal length. International J. of Poult. Sci., 2: 214-219.
- Zulkifli, I., N. Abdullah, N.M. Azrin and Y.W. Ho (2000). Broiler strains fed diets containing Lactobacillus cultures and oxytetracycline under heat stress conditions. Br.Poult. Sci., 41: 593-597.

تأثير اضافة منشطات النمو البيولوجية لعلائق بداري التسمين علي الإنتاجية والإستجابة المناعية وصفات الذبح سوزان أحمد رياض¹، حسام محمد صفاء¹، فاطمة رسمى محمد¹، سلوى سالم صيام² و حنان عبد العظيم المنشاوى³ ¹ قسم الانتاج الحيوانى – كلية الزراعة – جامعة القاهرة – جيزة – مصر ² قسم تربية الدواجن – معهد بحوث الانتاج الحيوانى – الدقى – جيزة – مصر ³ وزارة الزراعة – الدقى –جيزة – مصر

تم استخدام عدد 630 كتكوت أربور ايكرز لدراسة تأثير أضافة المنشطات البيولوجية في العلف على معدل الانتاج والإستجابة المناعية وخصائص الذبيحة. قُسمت الكتاكيت عند عمر يوم عشوائيا الى 6 مجموعات وكل مجموعة اشتملت علي 3 مكررات بكل مكرر 35 طائر، وكانت المجاميع كما يلى:

- المجموعة الأولى = الكنترول تغذت بها الكتاكيت على العليقة الاساسية بدون اضافات.
- 2- المجموعة الثانية = أَضِيف الى العليقة الاساسية مادة البروبيوتيك بمعدل 1جم/كجم علف.
- 3- المجموعة الثالثة = أضيف الى العليقة الاساسية مادة الخميرة يحتوي على 5×10 CFU 12 بمعدل 1جم/كجم علف.
 - 4- المجموعة الرابعة = أضيف الى العليقة الاساسية مادة البريبيوتيك بمعدل 1جم/كجم علف.

- 5- المجموعة الخامسة = أضيف الى العليقة الإساسية مادة البروبيوتيك + البريبيوتيك بمعدل 1جم لكل منهما/كجم علف.
- 6- المجموعة السادسة = أضيف الى العليقة الإساسية مادة الخميرة + البريبيوتيك بمعدل
 1 جم لكل منهما/كجم علف.

أظهرت النتائج أن استخدام الأضافات البيولوجية بالمعدلات المذكورة سالفاً فى العلف أدى الى تحسن وزن الجسم والزيادة الوزنية واستهلاك العلف والكفاءة التحويلية وصفات الذبح، كما أدي إلي خفض معدلات النفوق. كذلك أدت الإضافات البيولوجية لعلائق بداري التسمين الى زيادة عدد خلايا كرات الدم الحمراء والبيضاء بأنواعها المختلفة، كما أدت إلى زيادة مستوي الأجسام المناعية ضد كرات الدم الحمراء للأغنام في المناعة الأولية والثانوية كمؤشر لتحسن المناعة والإستجابة المناعية. أوضحت النتائج أيضاً أن أفضل معاملة هى المعاملة السادسة (أضافة الخميرة مع البريبيوتيك للعلف) كما أنها أعطت أعلى كفاءة اقتصادية. لذا توصي النتائج باضافة المنشطات البريبيولوجية لعلائق بداري التسمين لما لها من أثر إيجابي علي تحسين الإنتاجية والمناعة وصفات الذبح و كذلك الكفاءة الإقتصادية.

قام بتحكيم البحث

أ.د / أمينه عبد المطلب السروى
 أ.د / إبراهيم الوردانى السيد حسن

كلية الزراعة – جامعة المنصورة كلية الزراعة – جامعة عين شمس

		Ex	perimental treatm	nents			
Traits Control		Probiotic	Yeast Prebiotic		Probiotic + Prebiotic	Yeast + Prebiotic	
			Live body weight	(g)			
One day	44.26±0.05	44.31±0.02	44.37±0.02	44.37±0.01	44.33±0.01	44.38±0.04	
14 days	276.50 ^d ±4.96	285.58 ^{cd} ±1.94	297.09 ^{bc} ±3.34	300.83 ^b ±3.80	308.50 ^{ab} ±3.51	319.82 ^a ±4.39	
28 days	897.46 ^e ±8.79	964.20 ^d ±2.67	977.12 ^{cd} ±5.70	990.43 [°] ±3.23	1014.45 ^b ±6.86	1038.41 ^a ±5.91	
42 days	1779.94 ^e ±2.90	1956.36 ^d ±16.37	1981.26 ^{cd} ±15.01	2000.72 ^c ±10.20	2157.98 ^b ±5.59	2305.06 ^a ±10.20	
		E	Body weight gain	(g)			
0-14 days	231.90 ^d ±6.46	241.59 ^{cd} ±2.15	252.72 ^{bc} ±3.35	256.46 ^b ±3.79	264.16 ^{ab} ±3.51	275.34 ^a ±4.46	
15-28 days	619.89 ^d ±6.12	677.45 [°] ±1.74	679.18 ^c ±2.87	688.87 ^c ±1.29	705.38 ^b ±3.71	717.35 ^a ±4.48	
29-42 days	882.46 ^d ±3.63	994.81 [°] ±11.55	1005.71 ^c ±8.56	1012.80 ^c ±7.33	1145.31 ^b ±8.72	1266.50 ^a ±2.01	
0-42 days	1734.90 ^e ±5.07	1912.65 ^d ±14.98	1936.84 ^{cd} ±15.03	1956.27 ^c ±10.22	2113.63 ^b ±5.54	2260.68 ^a ±6.81	

Table (2): Live body weight (g) and body weight gain (g) of Arbor Acres broiler chicks fed diets with different biological additives from 0 to 42 days of age (Means±SEM).

a,b,c,d,e Means within rows with different superscripts are significantly different (P≤0.05).

Table (3): Feed consumption (g/bird/day) and feed conversion ratio (g feed/g gain) of Arbor Acres broiler chicks	
fed diets with different biological additives from 0 to 42 days of age (Means±SEM).	

	Experimental treatments									
Traits Age	Control Probiotic Yeast Prebiotic		Prebiotic	Probiotic + Prebiotic	Yeast + Prebiotic					
		Feed	consumption (g/c	hick/day)						
0-14 days	27.13 ^b ±1.01	29.34 ^a ±0.41	29.82 ^a ±0.44	29.36 ^a ±0.58	30.47 ^a ±0.39	30.72 ^a ±0.54				
15-28 days	85.97 ^b ±0.51	86.28 ^b ±0.45	87.98 ^{ab} ±0.31	87.69 ^{ab} ±1.25	88.39 ^{ab} ±1.00	90.14 ^a ±0.18				
29-42 days	112.60 ^d ±0.97	121.12 ^c ±3.82	131.00 ^b ±2.34	134.87 ^{ab} ±1.49	139.44 ^a ±1.36	141.04 ^a ±0.73				
0-42 days	77.79 ^c ±1.61	80.19 ^c ±1.37	84.42 ^b ±1.21	86.64 ^{ab} ±0.67	88.46 ^a ±0.97	88.24 ^a ±0.92				
		Feed	conversion (g fee	ed/g gain)						
0-14 days	1.63 ^{bc} ±0.09	1.70 ^a ±0.08	1.65 ^b ±0.07	1.60 ^c ±0.05	1.61 [°] ±0.09	1.56 ^d ±0.10				
15-28 days	1.94 ^a ±0.07	1.78 ^{bc} ±0.06	1.81 ^b ±0.09	1.78 ^{bc} ±0.07	1.75 [°] ±0.06	1.75 [°] ±0.08				
29-42 days	1.79 ^b ±0.13	1.70 ^c ±0.10	1.82 ^{ab} ±0.12	1.86 ^a ±0.08	1.70 ^c ±0.08	1.56 ^d ±0.09				
0-42 days	1.88 ^a ±0.08	1.76 ^c ±0.06	1.83 ^b ±0.04	1.86 ^{ab} ±0.03	1.75 ^c ±0.04	1.64 ^d ±0.05				

^{a,b,c,d,e} Means within rows with different superscripts are significantly different (P≤0.05).

Experimental treatments								
Traits Age	Control	Probiotic	Yeast	Prebiotic	Probiotic + Prebiotic	Yeast + Prebiotic		
0-7 days	1.63±1.25	0.00±0.00	0.00±0.00	0.00±0.00	0.00±0.00	0.54±0.54		
8-14 days	0.00±0.00	0.00±0.00	0.00±0.00	0.00±0.00	1.90±1.90	0.00±0.00		
15-21 days	6.13±3.17	1.93±0.97	2.03±1.09	1.63±1.44	1.03±0.94	1.37±1.37		
22-28 days	0.97±0.97	1.23±1.23	1.80±0.38	1.40±0.74	2.63±1.28	0.00±0.00		
29-35 days	1.27±1.27	0.70±0.70	0.83±0.83	2.83±1.52	2.10±1.72	0.00±0.00		
36-42 days	3.27±1.88	1.70±0.40	1.83±0.13	2.90±1.41	2.80±0.67	1.90±0.25		
0-42 days	13.27 ^a ±1.42	5.56 ^{bc} ±0.81	6.49 ^{bc} ±0.74	8.76 ^{abc} ±2.60	10.46 ^{ab} ±2.50	3.81 ^c ±1.78		

Table (4): Mortality rate (%) of Arbor Acres broiler chicks fed different biological additives in diets from 0 to 42 days of age.

^{a,b,c,d,e} Means within rows with different superscripts are significantly different (P≤0.05).

Table (5): Blood hematological picture and the antibody titer against SRB²C of Arbor Acres broiler chicks fed different biological additives from 0 to 42 days of age (Means±SEM).

Experimental treatments								
Traits Items	Control	Probiotic	Yeast	Prebiotic	Probiotic + Prebiotic	Yeast + Prebiotic		
	Blood he	ematological pi	cture at 42 day	s of age				
RBC's (x10 ⁶ /mm ³)	2.86 ^d ±0.01	3.23 ^c ±0.01	4.02 ^b ±0.01	4.26 ^b ±0.01	5.03 ^ª ±0.15	5.03 ^a ±0.01		
WBC's (x10 ³ /mm ³)	19.73 [†] ±0.07	23.34 ^e ±0.33	25.57 ^ª ±0.59	28.50 ^c ±0.10	31.06 ^b ±0.25	34.56 ^a ±0.44		
Lymphocyte (L; %)	51.02 ^b ±0.31	55.32 ^{ab} ±0.38	49.85 ^b ±0.72	59.27 ^{ab} ±0.40	61.37 ^{ab} ±0.60	65.32 ^a ±0.56		
Heterophils (H; %)	23.10 ^e ±0.21	25.32 ^d ±0.34	27.20 ^d ±0.42	30.68 ^c ±0.25	33.57 ^b ±0.34	37.14 ^a ±0.67		
H/L ratio	0.45±0.01	0.46±0.01	0.55±0.09	0.52±0.01	0.55±0.01	0.57±0.01		
Antibody titer against SRB'C								
Primary response at 35 days	41.40 ^c ±0.36	44.33 ^{bc} ±0.77	46.15 ^b ±1.35	47.26 ^b ±1.21	48.38 ^{ab} ±0.35	51.47 ^a ±0.64		
Secondary response at 42 days	183.12 ^e ±2.09	206.63 ^d ±1.15	243.28 ^b ±1.02	245.16 ^b ±0.29	234.26 [°] ±0.56	255.44 ^a ±0.47		

a,b,c,d,e,[†] Means within rows with different superscripts are significantly different (P≤0.05).

	Experimental treatments								
Traits Items	Control	Probiotic	Yeast	Prebiotic	Probiotic + Prebiotic	Yeast + Prebiotic			
Carcass (g)	971.11 ^e ±12.63	1126.67 ^d ±11.79	1156.67 ^d ±21.79	1211.11 ^c ±13.17	1331.11 ^b ±11.36	1464.44 ^a ±19.66			
Carcass (%)	54.83 ^e ±0.55	57.13 ^d ±0.33	58.17 ^{cd} ±0.55	59.26 ^c ±0.37	60.98 ^b ±0.35	62.33 ^a ±0.25			
Dressing (%)	59.20 ^e ±0.60	61.75 ^d ±0.36	62.82 ^{cd} ±0.57	63.98 ^c ±0.43	65.86 ^b ±0.45	67.37 ^a ±0.26			
Liver (%)	2.55 ^d ±0.04	2.63 ^{cd} ±0.02	2.65 ^{bcd} ±0.05	2.68 ^{bc} ±0.04	2.77 ^{ab} ±0.05	2.88 ^a ±0.03			
Heart (%)	0.40 ^b ±0.02	0.43 ^{ab} ±0.02	0.43 ^{ab} ±0.02	0.44 ^{ab} ±0.02	0.47 ^a ±0.03	0.48 ^a ±0.01			
Gizzard (%)	1.42 ^c ±0.03	1.56 ^b ±0.02	1.57 ^b ±0.02	1.60 ^b ±0.03	1.64 ^{ab} ±0.04	1.68 ^a ±0.02			
Total Giblets (%)	4.38 ^d ±0.08	4.62 ^c ±0.05	4.66 ^{bc} ±0.07	4.72 ^{bc} ±0.10	4.88 ^{ab} ±0.11	5.03 ^a ±0.06			
Abdominal fat (%)	1.69 ^a ±0.03	1.27 ^b ±0.02	1.24 ^b ±0.03	1.20 ^{bc} ±0.04	1.13 ^{cd} ±0.05	1.08 ^d ±0.03			
Edible part (%)	60.89 ^e ±0.62	63.03 ^d ±0.38	64.06 ^{cd} ±0.58	65.18 ^c ±0.46	66.98 ^b ±0.50	68.45 ^a ±0.28			
Intestine (%)	4.90±0.06	4.93±0.04	5.02±0.14	5.02±0.11	5.16±0.08	5.17±0.03			
Thymus (%)	0.18 ^c ±0.02	0.24 ^{bc} ±0.03	0.26 ^{abc} ±0.03	0.27 ^{ab} ±0.03	0.29 ^{ab} ±0.03	0.33 ^a ±0.02			
Bursa (%)	0.16 ^c ±0.02	0.17 ^{bc} ±0.02	0.18 ^b ±0.02	0.18 ^b ±0.02	0.21 ^a ±0.02	0.22 ^a ±0.01			
Spleen (%)	0.12 ^b ±0.02	0.14 ^{ab} ±0.02	0.14 ^{ab} ±0.02	0.14 ^{ab} ±0.02	0.16 ^{ab} ±0.01	0.19 ^a ±0.01			

Table (7): Slaughter traits at 42 days of age of Arbor Acres broiler chicks fed diets with supplemental different biological additives from 0 to 42 days of age (Means±SEM).

^{a,b,c,d,e} Means within rows with different superscripts are significantly different (P≤0.05).

Table (8): Economical efficiency of Arbor Acres broil	er chicks fed diets supplemented with different biological
additives at 42 days of age.	

Experimental treatments									
Items	Traits	Control	Probiotic	Yeast	Prebiotic	Probiotic + Prebiotic	Yeast + Prebiotic		
	Total intake (kg/chick)	3.27	3.37	3.55	3.64	3.72	3.71		
Feed	Price/kg (L.E)	2.06	2.10	2.07	2.09	2.19	2.16		
	Total feed cost (L.E)	6.73	7.07	7.34	7.61	8.14	8.01		
	Wight gain (kg/chick)	1.73	1.91	1.94	1.96	2.11	2.26		
Meat	Price/kg (L.E)	11.50	11.50	11.50	11.50	11.50	11.50		
	Total Revenue (L.E)	19.90	21.97	23.31	22.54	24.27	25.99		
Net Reven	ue (L.E)	13.17	14.89	15.97	14.93	16.13	17.98		
Economic	al efficiency	195.69	210.60	217.57	196.18	198.15	224.46		
Relative e	conomical efficiency (%)	100	107.62	111.18	100.25	101.26	114.70		

Net Revenue = Total Revenue - Total feed cost

Economic efficiency = Net Revenue / Total feed cost x 100

Relative economical efficiency (%) assuming the control treatment equal= 100%.